

ISSN 2312-9581 (Online)
ISSN 2075-1508 (Print)
DOI: <https://doi.org/10.61976/fsu>

F S U

FISHERIES SCIENCE OF UKRAINE

Institute of Fisheries
of the National Academy
of Agrarian Sciences
of Ukraine

Fisheries Science of Ukraine No. 1(71) – 2025

РИБОГОСПОДАРСЬКА НАУКА УКРАЇНИ

1 (71)

Інститут рибного господарства
Національної академії аграрних наук України

2025

РИБОГОСПОДАРСЬКА НАУКА УКРАЇНИ

Науковий журнал «Рибогосподарська наука України» з 2009 року входить до Переліку наукових фахових видань України, в яких можуть публікуватися результати дисертаційних робіт на здобуття ступенів доктора і кандидата наук, доктора філософії (галузі знань Н — «Сільське, лісове, рибне господарство та ветеринарна медицина» та Е — «Природничі науки, математика та статистика»), категорія «Б» (за спеціальностями 091 — Біологія, 207 — Водні біоресурси та аквакультура), переатестація від 17.03.2020 (наказ Міністерства освіти і науки України № 409)

Засновник та видавник — Інститут рибного господарства Національної академії аграрних наук України (ІРГ НААН)

Свідоцтво про державну реєстрацію КВ № 20934-10734 ПР від 18.08.2014

ГОЛОВНИЙ РЕДАКТОР — Грициняк І. І., д. с.-г. н., професор, академік НААН, в.о. директора ІРГ НААН, м. Київ, УКРАЇНА

ЗАСТУПНИКИ ГОЛОВНОГО РЕДАКТОРА (є штатними працівниками ІРГ НААН) — Матвієнко Н. М. (д. б. н., професор, зав. лаб. іхтіопатологія) та Симонов М. Ю. (к. с.-г. н., зав. лаб. науково-технічного співробітництва та інтелектуальної власності)

РЕДАКЦІЙНА КОЛЕГІЯ

Бузевич І. Ю., д. б. н., с. н. с., член-кор. НААН, зав. відділу дослідження біоресурсів водосховищ, ІРГ НААН, м. Київ, УКРАЇНА

Бучацький Л. П., д. б. н., проф., академік Академії наук Вищої школи України, пр. н. с. лаб. біотехнологій, ІРГ НААН, м. Київ, УКРАЇНА

Лисиця А. В., д. б. н., професор, ДС епізоотології, ННЦ «Інститут експериментальної і клінічної ветеринарної медицини», м. Рівне, УКРАЇНА

Новіцький Р. О., д. б. н., професор, зав. каф. водних біоресурсів та аквакультури, Дніпровський державний аграрно-економічний університет, м. Дніпро, УКРАЇНА

Потрохов О. С., д. б. н., с. н. с., зав. лаб. біології відтворення риб, Інститут гідробіології НАН України, м. Київ, УКРАЇНА

Рудик-Леуська Н. Я., д. б. н., доцент, зав. каф. гідробіології та іхтіології НУБіП України, м. Київ, УКРАЇНА

Сондак В. В., д. б. н., професор кафедри водних біоресурсів, Національний університет водного господарства та природокористування, м. Рівне, Україна

Діденко О. В., к. б. н., с. н. с., пр. н. с. відділу вивчення біоресурсів водосховищ, ІРГ НААН, м. Київ, УКРАЇНА

Забитівський Ю. М., к. б. н., ст. н. с. відділу популяційної екології Інституту екології Карпат НАНУ, м. Львів, УКРАЇНА;

Кружиліна С. В., к. б. н., с. н. с., пр. н. с. відділу вивчення біоресурсів водосховищ, ІРГ НААН, м. Київ, УКРАЇНА

Кутіщев П. С., к. б. н., доцент кафедри водних біоресурсів та аквакультури, Херсонський державний аграрний університет, м. Херсон, УКРАЇНА

Маренков О. М., к. б. н., доцент, проректор з наукової роботи, Дніпровський національний університет ім. Олеса Гончара, м. Дніпро, УКРАЇНА

Рудь Ю. П., к. б. н., с. н. с., зав. лаб. біотехнологій, ІРГ НААН, м. Київ, УКРАЇНА

Самчишина Л. В., к. б. н., пр. н. с. лаб. гідробіології та технологій культивування цінних безхребетних, ІРГ НААН, м. Київ, УКРАЇНА

Стачник М., PhD, відділ іхтіопатології, Національний дослідницький Інститут ветеринарії, м. Пулави, РЕСПУБЛІКА ПОЛЬЩА

Гриневич Н. Є., д. вет. н., професор, зав. каф. іхтіології та зоології, Білоцерківський національний аграрний університет, м. Біла Церква, УКРАЇНА

Христенко Д. С., к. б. н., с. н. с., ст. н. с. відділу вивчення біоресурсів водосховищ, ІРГ НААН, м. Київ, УКРАЇНА

Бех В. В., д. с.-г. н., професор, зав. кафедри аквакультури, Національний університет біоресурсів і природокористування, м. Київ, УКРАЇНА

Гламузіна Б., PhD, професор, кафедра аквакультури, Університет Дубровника, м. Дубровник, РЕСПУБЛІКА ХОРВАТІЯ

Димань Т. М., д. с.-г. н., професор, проректор з освітньої, виховної та міжнародної діяльності, Білоцерківський національний аграрний університет, м. Біла Церква, УКРАЇНА

Зодапе Г. В., д. н., професор, декан факультету зоології, Державний університет Шиваджи, м. Колхпур, РЕСПУБЛІКА ІНДІЯ

Капуста А., д. н., професор, Національний Дослідницький Інститут рибицтва у внутрішніх водах ім. Станіслава Саковича, м. Ольштин, РЕСПУБЛІКА ПОЛЬЩА

Пекарик Л., PhD, с. н. с., Центр рослинництва та біорізноманіття Словацької академії наук, м. Братислава, СЛОВАЦЬКА РЕСПУБЛІКА

Лобойко Ю. В., д. с.-г. н., доцент, зав. кафедри водних біоресурсів та аквакультури, Львівський національний університет ветеринарної медицини та біотехнологій ім. С. З. Гжицького, м. Львів, УКРАЇНА

Третяк О. М., д. с.-г. н., с. н. с., заступник директора з наукової роботи, ІРГ НААН, м. Київ, УКРАЇНА

Шек П. В., д. с.-г. н., професор кафедри водних біоресурсів та аквакультури, Одеський національний університет ім. І. І. Мечникова, м. Одеса, УКРАЇНА

Бреус Д. С., к. с.-г. н., доцент кафедри екології та сталого розвитку ім. професора Ю. В. Пилипенка, Херсонський державний аграрний університет, м. Херсон, УКРАЇНА

Ганкевич Б. О., к. с.-г. н., с. н. с. лаб. лососівництва, осетрівництва та технологій відтворення рідкісних та зникаючих видів риб, ІРГ НААН, м. Київ, УКРАЇНА

Костенко О. І., к. с.-г. н., с. н. с., заст. академіка-секретаря Відділення зоотехнії, Національна академія аграрних наук України, м. Київ, УКРАЇНА

Полікатор Т., PhD, професор, факультет рибицтва та захисту вод, Університет Південної Богемії, м. Чеські Будейовиці, ЧЕСЬКА РЕСПУБЛІКА

Чепіль Л. В., к. с.-г. н., доцент, кафедра біології тварин, Національний університет біоресурсів і природокористування, м. Київ, УКРАЇНА

Шарамок Т. С., к. с.-г. н., доцент, кафедра загальної біології та водних біоресурсів, Дніпровський національний університет імені Олеса Гончара, м. Дніпро, УКРАЇНА

НАУКОВІ РЕДАКТОРИ РОЗДІЛІВ (є штатними працівниками ІРГ НААН):

Бернакевич О. М., к. с.-г. н., с. н. с., в.о. директора Львівської дослідної станції ІРГ НААН; **Григоренко Т. В.**, к. с.-г. н., зав. лаб. гідробіології та технологій культивування цінних безхребетних; **Дерень О. В.**, к. с.-г. н., с. н. с., зав. лаб. кормів і годівлі риб;

Драган Л. П., к. б. н., с. н. с., зав. лаб. екологічних досліджень, **Куріненко Г. А.**, к. с.-г. н., ст. доцент, зав. лаб. селекції риб; **Кучерук А. І.**, к. с.-г. н., зав. лаб. лососівництва, осетрівництва та технологій відтворення рідкісних та зникаючих видів риб; **Маріуца А. Е.**, к. с.-г. н., с. н. с., зав. лаб. молекулярно-генетичних досліджень.

Редакція журналу «Рибогосподарська наука України»: вул. Обухівська, 135, м. Київ-164, 03164, тел.: +38 (098) 837-7150, <https://fsu.ua/>; e-mail: fsu.journal@gmail.com

Підписано до друку 18.03.2025 р. Формат 70×108/16. Друк офсетний. Ум. друк. арк. 8,0. Наклад 500 прим.

Журнал друкується за рішенням вченої ради Інституту рибного господарства НААН (Протокол № 3 від 18 березня 2025 року)

Дизайн макету: Архангельський Є. Ю., Симон М. Ю., Швець Т. М.

Верстка: Архангельський Є. Ю., Симон М. Ю. Літературне редагування: Швець Т. М. Коректор: Ковальчук Г. В.

Друкарня ТОВ «ПРО ФОРМАТ», 02166, м. Київ, вул. Кубанської України, 45 Б, оф. 16, тел.: +38(044) 353-85-58

DOI: [HTTPS://DOI.ORG/10.61976/FSU2025.01.137](https://doi.org/10.61976/FSU2025.01.137)*Ir. Hrytsyniak, T. Shvets***Crucian carp (*Carassius carassius* Linnaeus, 1758). Thematic bibliography***І. Й. Грициняк, Т. М. Швець***Карась золотий (Карась звичайний) (*Carassius carassius* Linnaeus, 1758).****Тематична бібліографія****137**DOI: [HTTPS://DOI.ORG/10.61976/FSU2025.01.163](https://doi.org/10.61976/FSU2025.01.163)*N. Hrynevych, A. Sliusarenko,**O. Khomiak, V. Zharchynska***Australian red-claw crayfish (*Cherax quadricarinatus* Von Martens, 1868).****Thematic bibliography***Н. Є. Гриневич, А. О. Слюсаренко,**О. А. Хом'як, В. С. Жарчинська***Австралійський червоноклешневий рак (*Cherax quadricarinatus* Von Martens, 1868).****Тематична бібліографія****163**

**AUSTRALIAN RED-
CLAW CRAYFISH (*CHERAX
QUADRICARINATUS* VON
MARTENS, 1868).
THEMATIC BIBLIOGRAPHY**

N. Hrynevych, gnatbc@ukr.net,
ORCID ID 0000-0001-7430-9498, Bila
Tserkva National Agrarian University, Bila
Tserkva

A. Sliusarenko, allasliusarenko@ukr.net,
ORCID ID 0000-0002-1896-8939, Bila
Tserkva National Agrarian University, Bila
Tserkva

O. Khomiak, chomiak_o@ukr.net,
ORCID ID 0000-0003-3010-6757, Bila
Tserkva National Agrarian University, Bila
Tserkva

V. Zharchynska, zharchynskavs@ukr.net,
ORCID ID 0000-0002-5823-9095, Bila
Tserkva National Agrarian University, Bila
Tserkva

Purpose. To compile scientific sources on the biology, ecology, reproduction and rearing of the Australian red-claw crayfish *Cherax quadricarinatus*, to identify the main areas of research related to the distribution, feeding and impact of this species on the ecosystem.

Methodology. The methodology for compiling the thematic bibliography was based on a thorough search, systematisation and analysis of scientific sources.

Findings. The work resulted in the collection, systemization, and analysis of a significant amount of literature covering the biological, ecological and economic aspects of the Australian red-claw crayfish *Cherax quadricarinatus*. The bibliography contains 154 publications, placed in alphabetical order and described in accordance with the requirements of the HAC, in accordance with DSTU 8302:2015 «Information and documentation. Bibliographic reference. General provisions and rules of drafting», taking into account the amendments (UKND code 01.140.40).

Practical Value. The list can be useful for students, teachers and researchers to access rel-

**АВСТРАЛІЙСЬКИЙ
ЧЕРВОНОКЛЕШНЕВИЙ РАК
(*CHERAX QUADRICARINATUS*
VON MARTENS, 1868).
ТЕМАТИЧНА БІБЛІОГРАФІЯ**

Н. Є. Гриневич, gnatbc@ukr.net,
ORCID ID 0000-0001-7430-9498,
Білоцерківський національний аграрний
університет, м. Біла Церква

А. О. Слюсаренко, allasliusarenko@ukr.net,
ORCID ID 0000-0002-1896-8939,
Білоцерківський національний аграрний
університет, м. Біла Церква

О. А. Хом'як, chomiak_o@ukr.net,
ORCID ID 0000-0003-3010-6757,
Білоцерківський національний аграрний
університет, м. Біла Церква

В. С. Жарчинська, zharchynskavs@ukr.net,
ORCID ID 0000-0002-5823-9095,
Білоцерківський національний аграрний
університет, м. Біла Церква

Мета. Упорядкування наукових джерел щодо біології, екології, особливостей відтворення та вирощування австралійського червоноклешневого рака *Cherax quadricarinatus*, виявлення основних напрямів наукових досліджень, що стосуються ареалу поширення, живлення та впливу цього виду на екосистему.

Методика. Методика складання тематичної бібліографії базувалася на ретельному пошуку, систематизації та аналізі наукових джерел.

Результати. У результаті проведеної роботи було зібрано, систематизовано та проаналізовано значний масив літературних джерел, що висвітлюють біологічні, екологічні та господарські характеристики австралійського червоноклешневого рака *Cherax quadricarinatus*. Бібліографія містить 154 публікації, розміщені у алфавітному порядку та описані згідно з вимогами ВАК, відповідно до ДСТУ 8302:2015 «Інформація та документація. Бібліографічне посилання. Загальні положення та правила складання», із урахуванням поправок (код УКНД 01.140.40).



evant publications for use in the educational process and research.

Keywords: aquaculture, *Cherax quadricarinatus*, distribution area, reproduction, ecdysis, morphometric analysis.

Практична значимість. Перелік може бути корисним для студентів, викладачів і науковців доступом до актуальних публікацій для використання в навчальному процесі та наукових роботах.

Ключові слова: аквакультура, *Cherax quadricarinatus*, ареал поширення, відтворення, екдизис, морфометричний аналіз.

REFERENCES

1. Abehsera, S., Bentov, S., Li, X., Weil, S., Manor, R., Sagi, S., Li, S., Li, F., Khalaila, I., Aflalo, E. D., & Sagi, A. (2021). Genes encoding putative bicarbonate transporters as a missing molecular link between molt and mineralization in crustaceans. *Scientific Reports*, 11(1), 11722. <https://doi.org/10.1038/s41598-021-91155-w>.
2. Abehsera, S., Zaccai, S., Mittelman, B., Glazer, L., Weil, S., Khalaila, I., Bitton, R., Zarivach, R., Li, S., Li, F., Xiang, J., Manor, R., Aflalo, E.D., & Sagi, A. (2018). CPAP3 proteins in the mineralized cuticle of a decapod crustacean. *Scientific Reports*, 8(1), 2430. <https://doi.org/10.1038/s41598-018-20835-x>
3. Abizar, Purnamasari, L., Widyawati, Affandi, M., & Putranto, T. W. C. (2020). Morphometric characteristics of crayfish *Cherax quadricarinatus* from atokan river, West Sumatera, Indonesia. *Ecology, Environment and Conservation*, 26(4), 1787–1792.
4. Akmal, S. G., Santoso, A., Yonvitner, Yuliana, E., & Patoka, J. (2021). Red-claw crayfish (*Cherax quadricarinatus*): spatial distribution and dispersal pattern in Java, Indonesia. *Knowledge and Management of Aquatic Ecosystems*, 422, 16. <https://doi.org/10.1051/kmae/2021015>.
5. Ali, M. Y., Pavasovic, A., Mather, P. B., & Prentis, P. J. (2017). Expression patterns of two carbonic anhydrase genes, Na⁺/K⁺-ATPase and V-type H⁺-ATPase, in the freshwater crayfish, *Cherax quadricarinatus*, exposed to low pH and high pH. *Australian Journal of Zoology*, 65(1), 50–59. <https://doi.org/10.1071/ZO16048>.

ЛІТЕРАТУРА

1. A comparative transcriptomic analysis in late embryogenesis of the red claw crayfish *Cherax quadricarinatus* / Wang Y. et al. // Molecular genetics and genomics. 2020. Vol. 295(2). P. 299—311. <https://doi.org/10.1007/s00438-019-01621-4>.
2. A crustacean annotated transcriptome (CAT) database / Nong W. et al. // BMC Genomics. 2020. Vol. 21(1). P. 32. <https://doi.org/10.1186/s12864-019-6433-3>.
3. A giant genome for a giant crayfish (*Cherax quadricarinatus*) with insights into cox1 pseudogenes in decapod genomes / Tan M. H. et al. // Frontiers in genetics. 2020. Vol. 11. P. 201. <https://doi.org/10.3389/fgene.2020.00201>
4. A review of freshwater crayfish introductions in Africa / Madzivanzira T. C. et al. // Reviews in Fisheries Science & Aquaculture. 2020. Vol. 29(2). P. 218—241. <https://doi.org/10.1080/23308249.2020.1802405>.
5. A study on the effects of light intensity on juveniles of the red claw crayfish *Cherax quadricarinatus* / Wen-ping J. et al. // Aquaculture Research. 2022. Vol. 53(18). P. 6454—6462. <https://doi.org/10.1111/are.16115>.
6. A transporter that allows phosphate ions to control the polymorph of exoskeletal calcium carbonate biomineralization / Shaked S. A. et al. // Acta Biomaterialia. 2024. Vol. 178. P. 221—232. <https://doi.org/10.1016/j.actbio.2024.02.035>.



6. Andriyeni, A., Zulkhasyni, Z., Athybi, G. S., & Pardiansyah, D. (2022). Effect of cutting organs of lobster shrimp (*Cherax quadricarinatus*) on moulting percentage and survival. *Jurnal Agroqua: Media Informasi Agronomi Dan Budidaya Perairan*, 20(1), 157–164. <https://doi.org/10.32663/ja.v20i1.2653>.
7. Arias, A., & Torralba-Burrial, A. (2021). First record of the redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) on the Iberian Peninsula. *Limnetica*, 40, 33–42. <https://doi.org/10.23818/limn.40.03>.
8. Azoifeifa-Solano, J. C., Carranza, A. H. R., Naranjo-Elizondo, B., & Fonseca, M. C. (2017). Presence of the Australian redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) (*Parastacidae*, *Astacoidea*) in a freshwater system in the Caribbean drainage of Costa Rica. *BioInvasions Records*, 6(4), 351–355. <https://doi.org/10.3391/bir.2017.6.4.08>.
9. Azoifeifa-Solano, J. C., Villalobos-Rojas, F., Romero-Chaves, R., & Wehrtmann, I. S. (2023). Modeling the habitat suitability of two exotic freshwater crayfishes in Mesoamerica and the Caribbean: *Cherax quadricarinatus* (von Martens, 1868) and *Procambarus clarkii* Girard, 1852 (*Decapoda: Astacidea: Parastacidae, Cambaridae*). *Journal of Crustacean Biology*, 43(4), ruad059. <https://doi.org/10.1093/jcabi/ruad059>.
10. Baudry, T., Becking, T., Goût, J. P., Arqué, A., Gan, H. M., Austin, C. M., Delaunay C., Juliette Smith-Ravin J., Roques J. A. C., & Grandjean, F. (2020). Invasion and distribution of the redclaw crayfish, *Cherax quadricarinatus*, in Martinique. *Knowledge and Management of Aquatic Ecosystems*, 421, 50. <https://doi.org/10.1051/kmae/2020041>.
11. Baudry, T., Gismondi, E., Goût, J. P., Arqué, A., Smith-Ravin, J., & Grandjean, F. (2022). The invasive crayfish *Cherax quadricarinatus* facing chlordecone in Martinique: bioaccumulation
7. Abizar Purnamasari L., Widayawati Af-fandi M., Putranto T. W. C. Morphometric characteristics of crayfish *Cherax quadricarinatus* from atokan river, West Sumatera, Indonesia // Ecology, Environment and Conservation. 2020. Vol. 26(4). P. 1787—1792.
8. Acute toxicity of cypermethrin on the juvenile of red claw crayfish *Cherax quadricarinatus* / Yuan J. et al. // Chemosphere. 2019. Vol. 237. 124468. <https://doi.org/10.1016/j.chemosphere.2019.124468>.
9. Additional records of wild populations of the Australian crayfish *Cherax quadricarinatus* in Mexico / Rodríguez-Almaraz G. A. et al. // Revista Mexicana de Biodiversidad. 2018. Vol. 89(4). P. 1322—1327. <https://doi.org/10.22201/ib.20078706e.2018.4.2065>.
10. Allergen diversity and abundance in different tissues of the redclaw crayfish (*Cherax quadricarinatus*) / Jerry E. M. et al. // Foods. 2024. Vol. 13(2). P. 315. <https://doi.org/10.3390/foods13020315>.
11. Also in crayfish: how phytase inclusion avoids phytic acid effects on hepatopancreas enzymes of redclaw *Cherax quadricarinatus* / Casaretto M. E. et al. // Aquaculture Research. 2023. Vol. 10. 4341218. <https://doi.org/10.1155/2023/4341218>.
12. An examination of the effectiveness of traps and baits as a possible means of harvesting crayfish, *Cherax quadricarinatus* in Sanyati Basin, Lake Kariba, Zimbabwe / Mhlanga L. et al. // Water SA. 2020. Vol. 46(4). P. 675—678. <https://doi.org/10.17159/wsa/2020.v46.i4.9083>.
13. An update of the known distribution and status of *Cherax* spp. in Italy (Crustacea, Parastacidae) / Vecchioni L. et al. // BioInvasions Records. 2022. Vol. 11(4).



- and depuration study. *Chemosphere*, 286, 131926. <https://doi.org/10.1016/j.chemosphere.2021.131926>.
12. Beatty, S. J., Ramsay, A., Pinder, A. M., & Morgan, D. L. (2020). Reservoirs act as footholds for an invasive freshwater crayfish. *Pacific Conservation Biology*, 26(1), 78–83. <https://doi.org/10.1071/PC19012>.
 13. Bian, Y., Liu, S., Liu, Y., Jia, Y., Li, F., Chi, M., Zheng J., Cheng S., & Gu, Z. (2022). Development of a multiplex PCR assay for parentage assignment of the red-claw crayfish (*Cherax quadricarinatus*). *Aquaculture*, 550, 737813. <https://doi.org/10.1016/j.aquaculture.2021.737813>.
 14. Cai, L., Zheng, J., Jia, Y., Gu, Z., Liu, S., Chi, M., & Cheng, S. (2020). Molecular characterization and expression profiling of three transformer-2 splice isoforms in the redclaw crayfish, *Cherax quadricarinatus*. *Frontiers in Physiology*, 11(631), 10. <https://doi.org/10.3389/fphys.2020.00631>.
 15. Calvo, N. S., Stumpf, L., Cortés-Jacinto, E., Castillo Díaz, F., & López Greco, L. S. (2018). Mobilization of energetic reserves during starvation in juveniles of different size of the redclaw crayfish *Cherax quadricarinatus*. *Aquaculture Nutrition*, 24(3), 952–960. <https://doi.org/10.1111/anu.12631>.
 16. Casaretto, M. E., Stumpf, L., Azcuy, R. L., López Greco, L. S., & Colombatto, D. (2023). Also in crayfish: how phytase inclusion avoids phytic acid effects on hepatopancreas enzymes of redclaw *Cherax quadricarinatus*. *Aquaculture Research*, 10, 4341218. <https://doi.org/10.1155/2023/4341218>.
 17. Chakandinakira, A. T., Madzivanzira, T. C., Mashonga, S., Muzvondiwa, J. V., Ndlovu, N., & South, J. (2023). Socioeconomic impacts of Australian redclaw crayfish *Cherax quadricarinatus* in Lake Kariba. *Biological Invasions*, 25(9), 2801–2812. <https://doi.org/10.1007/s10530-023-03074-8>.
 18. Chen, H., Zhang, R., Liu, F., Shao, C., Liu, F., Li, W., Ren, J., Niu, B., Liu, H., & Lou, B. (2023). The chromosome-level P. 1045—1055. <https://doi.org/10.3391/bir.2022.11.4.22>.
 14. Analysis of change of nutrients from fertilized eggs to newly hatched shrimp of *Cherax quadricarinatus* / Sun L. et al. // Journal of Shanghai Ocean University. 2023. Vol. 32(2). P. 348—356. <https://doi.org/10.12024/jsou.20220203718>.
 15. Analysis of the differences in muscle nutrition among individuals of different sexes in redclaw crayfish, *Cherax quadricarinatus* / Sun Y. et al. // Metabolites. 2023. Vol. 13(2). P. 190. <https://doi.org/10.3390/metabo13020190>.
 16. Analysis of transcripts and splice isoforms in red claw crayfish (*Cherax quadricarinatus*) using single-molecule long-read sequencing / Xu Y. et al. // Aquaculture. 2021. Vol. 541. 736828. <https://doi.org/10.1016/j.aquaculture.2021.736828>.
 17. Analysis on phenotypic traits and muscle nutritional composition of *Cherax quadricarinatus* in different specifications / Sun L. H. et al. // Oceanologia et Limnologia Sinica. 2023. Vol. 54(3). P. 885—894. <https://doi.org/10.11693/hyhz20220800220>.
 18. Antibacterial activity of freshwater lobster (*Cherax quadricarinatus*) shell chitosan gel preparation against *Escherichia coli* and *Staphylococcus aureus* / Rani Z. et al. // Journal of Applied Pharmaceutical Science. 2023. Vol. 13(2). P. 146—153. <https://doi.org/10.7324/JAPS.2023.130216>.
 19. Arias A., Torralba-Burrial A. First record of the redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) on the Iberian Peninsula // Limnetica. 2021. Vol. 40. P. 33—42. <https://doi.org/10.23818/limn.40.03>.
 20. Asymmetric firing rate from crayfish left



- el genome of *Cherax quadricarinatus*. *Scientific data*, 10(1), 215. <https://doi.org/10.1038/s41597-023-02124-z>.
19. Chen, H., Zhang, R., Liu, F., Shao, C., Liu, F., Li, W., Ren, J., Niu, B., Liu, H., & Lou, B. (2023). Publisher Correction: The chromosome-level genome of *Cherax quadricarinatus*. *Scientific Data*, 10(1), 313. <https://doi.org/10.1038/s41597-023-02186-z>.
 20. Chen, L., Zheng, J., Jia, Y., Li, F., Gu, Z., Chi, M., Cheng, S., Liu, S., Jiang, W., & Liu, Y. (2022). Molecular characterization of the Ftz-f1 gene in redclaw crayfish *Cherax quadricarinatus* and its potential role in ovarian development. *Aquaculture Research*, 53(15), 5261–5269. <https://doi.org/10.1111/are.16010>.
 21. Chen, Q., Lv, W., Jiao, Y., Liu, Z., Li, Y., Cai, M., Wu, D., Zhou, W., & Zhao, Y. (2020). Effects of exposure to waterborne polystyrene microspheres on lipid metabolism in the hepatopancreas of juvenile redclaw crayfish, *Cherax quadricarinatus*. *Aquatic toxicology*, 224, 105497. <https://doi.org/10.1016/j.aquatox.2020.105497>.
 22. Cheng, S., Zheng, J.-B., Jia, Y.-Y., Chi, M.-L., Jiang, W.-P., Liu, S.-L., Li, F., Liu, Y.-N., Gu, Z.-M., & Wang, D.-L. (2023). Effects of light color, photoperiod, and growth-related gene interference or overexpression on the survival, growth, or physiological and biochemical indices of red claw crayfish juveniles. *Aquaculture*, 562, 738740. <https://doi.org/10.1016/j.aquaculture.2022.738740>.
 23. Chivambo, S., Mussagy, A., & Barki, A. (2020). Assessment of interspecific interactions between the invasive red-claw crayfish (*Cherax quadricarinatus*) and the mozambique tilapia (*Oreochromis mossambicus*). *Brazilian Journal of Biology*, 80(4), 717–726. <https://doi.org/10.1590/1519-6984.217868>.
 24. Crandall, K. A., & Grave, S. D. (2017). An updated classification of the freshwater crayfishes (*Decapoda: Astacidea*) of the and right caudal photoreceptors due to blue and green monochromatic light pulses / Sánchez-Hernández J. C. et al. // *Symmetry*. 2018. Vol. 10(9). P. 389. <https://doi.org/10.3390/sym10090389>.
 21. cDNA characterization and expression of selenium-dependent CqGPx3 isoforms in the crayfish *Cherax quadricarinatus* under high temperature and hypoxia / Hernández-Aguirre L. E. et al. // *Genes*. 2022. Vol. 13(2). P. 179. <https://doi.org/10.3390/genes13020179>.
 22. Chivambo S., Mussagy A., Barki A. Assessment of interspecific interactions between the invasive red-claw crayfish (*Cherax quadricarinatus*) and the Mozambique tilapia (*Oreochromis mossambicus*) // *Brazilian Journal of Biology*. 2020. Vol. 80(4). P. 717–726. <https://doi.org/10.1590/1519-6984.217868>.
 23. Cloning and characterisation of a $\Delta 9$ fatty acyl desaturase-like gene from the red claw crayfish (*Cherax quadricarinatus*) and its expression analysis under cold stress / Wu D. L. et al. // *Journal of Thermal Biology*. 2021. Vol. 102. 103122. <https://doi.org/10.1016/j.jtherbio.2021.103122>.
 24. Clove oil as an anaesthetic for Australian redclaw crayfish *Cherax quadricarinatus* / Ghanawi J. et al. // *Aquaculture Research*. 2019. Vol. 50(12). P. 3628–3632. <https://doi.org/10.1111/are.14319>.
 25. Cold stress regulates lipid metabolism via AMPK signalling in *Cherax quadricarinatus* / Wu D. et al. // *Journal of thermal biology*. 2020. Vol. 92. 102693. <https://doi.org/10.1016/j.jtherbio.2020.102693>.
 26. Color quality improvement of *Cherax quadricarinatus* with dragon fruit peel meal utilization / Rakhmawati R. et al. // *Depik*. 2023. Vol. 12(3). P.



- world, with a complete species list. *Journal of Crustacean Biology*, 37(5), 615–653. <https://doi.org/10.1093/jcbiol/rux070>.
25. Cui, Y., Hao, G., Lin, F., Zhou, D., Sheng, P., Ding, L., & Zhang, H. (2020). Comparison of the nutrition and texture of *Cherax quadricarinatus* muscle in two culture patterns. *Food and Fermentation Industries*, 46(21), 115–120. <https://doi.org/10.13995/j.cnki.11-1802/ts.023977>.
26. Dai, P., Zheng, J., Luan, S., Kong, J., Jia, Y., & Gu, Z. (2022). Estimates of heritability and genetic correlation for growth traits at harvest in redclaw crayfish, *Cherax quadricarinatus*. *Aquaculture*, 561, 738631. <https://doi.org/10.1016/j.aquaculture.2022.738631>.
27. Dali, M. Z. M., Nasir, M. S. A. M., Khaleel, A. G., Chun, L. M., Gan, H. M., Wan, N. A. F. N., Umar, R., & Kamarudin, A. S. (2023). Predicting *Cherax quadricarinatus* habitat distribution patterns through the usage of GIS and eDNA analysis in Terengganu, Malaysia. *Sains Malaysiana*, 52(2), 343–354. <https://doi.org/10.17576/jism-2023-5202-03>.
28. Dali, M. Z. M., Nasir, M. S. A. M., Khaleel, A. G., Madiran, N. A., Ismail, N., & Kamarudin, A. S. (2023). Genetic variability of wild populations of invasive redclaw crayfish (*Cherax quadricarinatus*) von Martens 1868 across peninsular Malaysia. *Malaysian Applied Biology*, 52(1), 35–42. <https://doi.org/10.55230/mabjournal.v52i1.2427>.
29. Daubnerová, I., & Žitňan, D. (2021). *Ecdysis triggering hormone*. *Handbook of Hormones (2nd edn.)*, 2, 829–831. <https://doi.org/10.1016/B978-0-12-820649-2.00224-2>.
30. Douthwaite, R. J., Jones, E. W., Tyser, A. B., & Vrdoljak, S. M. (2018). The introduction, spread and ecology of redclaw crayfish *Cherax quadricarinatus* in the Zambezi catchment. *African Journal of Aquatic Science*, 43, 353–366. <https://doi.org/10.2989/16085914.2018.1517080>.
- 314–319. <https://doi.org/10.13170/de-pik.12.3.28628>.
27. Comparative study on the nutritional content and physical attributes of giant freshwater prawn (*Macrobrachium rosenbergii*) and redclaw crayfish (*Cherax quadricarinatus*) meats: preprint (Version 1) 07 June / Zheng-Bin T. et al. // Research Square. <https://doi.org/10.21203/rs.3.rs-1695209/v1>.
28. Comparison of growth performance, carotenoid content, and temperature tolerance of two-colored strains of the red claw crayfish *Cherax quadricarinatus* / Wei M. et al. // Journal of Shellfish Research. 2021. Vol. 40(2). P. 421–427. <https://doi.org/10.2983/035.040.0214>.
29. Comparison of the nutrition and texture of *Cherax quadricarinatus* muscle in two culture patterns / Cui Y. et al. // Food and Fermentation Industries. 2020. Vol. 46(21). P. 115–120. <https://doi.org/10.13995/j.cnki.11-1802/ts.023977>.
30. CPAP3 proteins in the mineralized cuticle of a decapod crustacean / Abehsera S. et al. // Scientific Reports. 2018. Vol. 8(1). 2430. <https://doi.org/10.1038/s41598-018-20835-x>.
31. Crandall K. A., Grave S. D. An updated classification of the freshwater crayfishes (Decapoda: Astacidea) of the world, with a complete species list. *Journal of Crustacean Biology*. 2017. Vol. 37(5). P. 615–653. <https://doi.org/10.1093/jcbiol/rux070>.
32. Crayfish hemocyanin on chitin bone substitute scaffolds promotes the proliferation and osteogenic differentiation of human mesenchymal stem cells / Krupke B. et al. // Journal of Biomedical Materials Research — Part A. 2020. Vol. 108(3). P. 694–708. <https://doi.org/10.1002/jbm.a.36849>.



31. Duan, H., Jin, S., Li, F., Zhang, X., & Xiang, J. (2018). Neuroanatomy and morphological diversity of brain cells from adult crayfish *Cherax quadricarinatus*. *Journal of Oceanology and Limnology*, 36(6), 2368–2378. <https://doi.org/10.1007/s00343-019-7145-x>.
32. Fahrudin, M., Suriyadin, A., & Murtawan, H. (2022). Pertumbuhan dan kelangsungan hidup lobster air tawar (*Cherax quadricarinatus*) dengan pemberian substrat yang berbeda. *Jurnal Marikultur*, 4(1), 31–41.
33. Fasya, A. H. (2019). Study of patterns in the relationship of ecdysis with the age of freshwater crayfish *Cherax quadricarinatus* aged 76 days. *IOP Conference Series: Earth and Environmental Science*, 236(1), 012012. <https://doi.org/10.1088/1755-1315/236/1/012012>.
34. Fatihah, S. N., Lim, L-S., Harman, M-F., & Ikhwanuddin, M. (2020). Effect of substrate on growth, survival and moulting in juvenile red claw, *Cherax quadricarinatus*. *Journal of PeerScientist*, 3(2), e1000027.
35. Fauzan Isma, M., & Syahril, M. (2021). Effects of difference shelter on survival rate and growth of freshwater lobster (*Cherax quadricarinatus*). *Jurnal Ilmiah Samudra Akuatika*, 5(1), 1–8. <https://doi.org/10.33059/jisa.v5i1.3547>.
36. Fedorovych, Ye. I., Muzhenko, A. V., & Sliusar M. V. (2022). Zviazok khimichnykh ta fizychnykh pokaznykiv vody z morfolohichnymy oznakamy rakiv riznykh vydiviu *Visnyk Sums-koho natsionalnoho ahrarnoho universytetu*, 4(47), 165–170. <https://doi.org/10.32845/bsnau.lvst.2021.4.28>.
37. Fedorovych, Ye. I., Muzhenko, A. V., Sliusar, M. V., & Kovalchuk, I. I. (2022). Osoblyvosti protsesu lynky rakiv riznykh vydiv. *Tavriiskyi naukovyi visnyk*, 126, 230–237. <https://doi.org/10.32851/2226-0099.2022.126.32>.
38. Fu, R., Li, F., & Yang, F. (2019). Separation of hemocytes of *Cherax quadricarinatus* by percoll discontinuous density
33. CrustyBase: an interactive online database for crustacean transcriptomes / Hyde C. J. et al. // *BMC Genomics*. 2020. Vol. 21. P. 637. <https://doi.org/10.1186/s12864-020-07063-2>.
34. Daubnerová I., Žitňan D. Ecdysis triggering hormone. *Handbook of Hormones*. 2nd edn. 2021. Vol. 2. P. 829–831. <https://doi.org/10.1016/B978-0-12-820649-2.00224-2>.
35. Development of a multiplex PCR assay for parentage assignment of the redclaw crayfish (*Cherax quadricarinatus*) / Bian Y. et al. // *Aquaculture*. 2022. Vol. 550. 737813. <https://doi.org/10.1016/j.aquaculture.2021.737813>.
36. Development of a primary culture system for haematopoietic tissue cells from *Cherax quadricarinatus* and an exploration of transfection methods / Xu X. et al. // *Developmental and Comparative Immunology*. 2018. Vol. 88. P. 45–54. <https://doi.org/10.1016/j.dci.2018.07.006>.
37. Development of economic valuation method for the direct impact of alien invasive species based on food competition in aquatic ecosystems / Kodiran T. et al. // *Jurnal Pengelolaan Sumberdaya Alam Dan Lingkungan*. 2020. Vol. 10(2). P. 198–208. <https://doi.org/10.29244/jpsl.10.2.198-208>.
38. Development of species-specific primer sets for Australian redclaw crayfish (*Cherax quadricarinatus*) detection from water environmental DNA (eDNA) / Nasir S. A. M. et al. // *Bioscience Research*. 2020. Vol. 17. P. 90–99.
39. Diet composition changes in tigerfish of Lake Kariba following an invasion by redclaw crayfish / Marufu L. et al. // *Annales de Limnologie*. 2017. Vol. 53. P. 47–56. <https://doi.org/10.1051/>



- gradient centrifugation. *Journal of Fisheries of China*, 43(4), 841–851. <https://doi.org/10.11964/jfc.20180511286>.
39. Ghanawi, J., Saoud, G., Zakher, C., Monzer, S., & Saoud, I.P. (2019). Clove oil as an anaesthetic for Australian red-claw crayfish *Cherax quadricarinatus*. *Aquaculture Research*, 50(12), 3628–3632. <https://doi.org/10.1111/are.14319>.
40. Hassan, M., Azri-Shah, N. N., Zakariah, M. I., Yusoff, N. A. H., Abdullah, F., Wahab, W., Ishak, A. N., Husin, N. M., & Jones, J. B. (2022). Morphological and molecular identification of *Dicreratocephala boschmai* Baer, 1953 and *Decadidymus* sp. Cannon, 1991 on wild and cultured environment of *Cherax quadricarinatus* in Malaysia. *BioInvasions Records*, 11(2), 495–509. <https://doi.org/10.3391/bir.2022.11.2.22>.
41. Hassan, M., Azri-Shah, N., Zakariah, M., Yusoff, N., Abdullah, F., Wahab, W., Ishak, A., Husin, N., & Jones, J. (2023). Prevalence of temnocephalids on cultured and wild *Cherax quadricarinatus* in Malaysia. *Egyptian Journal of Aquatic Research*, 49(3), 395–400. <https://dx.doi.org/10.1016/j.ejar.2022.03.004>.
42. Hayakijkosol, O., Jaroenram, W., Owens, L., & Elliman, J. (2021). Reverse transcription polymerase chain reaction (RT-PCR) detection for Australian *Cherax reovirus* from redclaw crayfish (*Cherax quadricarinatus*). *Aquaculture*, 530, 735881. <https://doi.org/10.1016/j.aquaculture.2020.735881>.
43. Hernández-Aguirre, L. E., Fuentes-Sidas, Y. I., Rivera-Rangel, L. R., Gutiérrez-Méndez, N., Yepiz-Plascencia, G., Chávez-Flores, D., Zavala-Díaz de la Serna, F.J., Peralta-Pérez, M. D. R., & García-Triana, A. (2022). cDNA characterization and expression of selenium-dependent CqGPx3 isoforms in the crayfish *Cherax quadricarinatus* under high temperature and hypoxia. *Genes*, 13(2), 179. <https://doi.org/10.3390/genes13020179>.
- limn/2016033.
40. Do opposites attack? Resource interactions between an alien and native crayfish from the lake Eyre basin / King G. et al. // Marine and Freshwater Research. 2022. Vol. 73(7). P. 873—883. <https://doi.org/10.1071/MF21302>.
41. Ecdysis triggering hormone modulates molt behaviour in the redclaw crayfish *Cherax quadricarinatus*, providing a mechanistic evidence for conserved function in molt regulation across Pancrustacea / Minh Nhut T. et al. // General and Comparative Endocrinology. 2020. Vol. 298(1). 113556. <https://doi.org/10.1016/j.ygcen.2020.113556>.
42. Effect of cutting organs of lobster shrimp (*Cherax quadricarinatus*) on moulting percentage and survival / Andriyeni A. et al. // Jurnal Agroqua: Media Informati Agronomi Dan Budidaya Perairan. 2022. Vol. 20(1). P. 157—164. <https://doi.org/10.32663/ja.v20i1.2653>.
43. Effect of feed additives in the diet on the growth and testicular development of male red claw crayfish (*Cherax quadricarinatus*) using orthogonal experiments / Shehata A. I. et al. // Animal Feed Science and Technology. 2022. Vol. 283. 115180. <https://doi.org/10.1016/j.anifeedsci.2021.115180>.
44. Effect of high hydrostatic pressure processing on biochemical characteristics, bacterial counts, and color of the red claw crayfish *Cherax quadricarinatus* / Lin X. et al. // Journal of Shellfish Research. 2021. Vol. 40. P. 177—184. <https://doi.org/10.2983/035.040.0117>.
45. Effect of substrate on growth, survival and moulting in juvenile red claw, *Cherax quadricarinatus* / Fatihah S. N. et al. // Journal of PeerScientist. 2020. Vol. 3(2). e1000027.



44. Hernández-Rubio, M. C., de Jesús Gutiérrez-Ladrón de Guevara, M., & Figueroa-Lucero, G. (2021). Shelters presence effect on development of early juvenile *Cherax quadricarinatus* (Decapoda; Parastacidae). *Hidrobiologica*, 30(3), 69–75. <https://doi.org/10.24275/uam/izt/dcbs/hidro/2021v31n1/Hernandez>.
45. Hou, Y., Jia, R., Sun, W., Ding, H., Li, B., & Zhu, J. (2023). Red claw crayfish *Cherax quadricarinatus* cultivation influences the dynamics and assembly of benthic bacterial communities in paddy fields. *Environments*, 10(10), 178. <https://doi.org/10.3390/environments10100178>.
46. Hrynevych, N. Ye., & Zharchynska, V. S. (2022). Osoblyvosti zovnishnoi budovy *Cherax quadricarinatus* (Von Martens, 1868). *Problems of science and practice, tasks and ways to solve them : XX International scientific and practical conference*. Warsaw, 44–46.
47. Hyde, C. J., Fitzgibbon, Q. P., Elizur, A., Smith, G. G., & Ventura, T. (2020). CrustyBase: an interactive online database for crustacean transcriptomes. *BMC Genomics*, 21, 637. <https://doi.org/10.1186/s12864-020-07063-2>.
48. Iqbal, M. A., Setyobudiandi, I., Krisanti, M., & Wardiatno, Y. (2019). Produksi telur *Cherax quadricarinatus* (Von Martens, 1868) di danau Lido, Jawa Barat. *Jurnal Pengelolaan Perikanan Tropis*, 3(2), 45–52.
49. Ismail, N., Nasir, M. S. A. M., Khaleel, A. G., Sallehuddin, A. S., Idrus, S. N. S., Istiqomah, I., Balu Alagar Venmathi Maran B. A. V., & Kamarudin, A. S. (2021). First wild record of australian redclaw crayfish *Cherax quadricarinatus* (Von Martens, 1868) in the east coast of peninsular Malaysia. *BioInvasions Records*, 10(2), 360–368. <https://doi.org/10.3391/bir.2021.10.2.14>.
50. Jaroenram, W., Hayakijkosol, O., Owens, L., & Elliman, J. (2021). Establishing a gold standard method for the detection of *Cherax reovirus* using reverse transcriptase, quantitative, polymerase
46. Effects of background color on growth, survival, body color, and inhabiting behavior distribution of *Cherax quadricarinatus* / Wei M. et al. // *Journal of Marine Sciences*. 2020. Vol. 44. P. 60—65.
47. Effects of eggshell and seashell powder as natural dietary calcium supplements on growth, molting frequency, and carapace calcium composition of juvenile red claw crayfish, *Cherax quadricarinatus* / Shahroom A. et al. // *PeerJ*. 2023. Vol. 11. e15449. <https://doi.org/10.7717/peerj.15449>.
48. Effects of exposure to waterborne polystyrene microspheres on lipid metabolism in the hepatopancreas of juvenile redclaw crayfish, *Cherax quadricarinatus* / Chen Q. et al. // *Aquatic toxicology*. 2020. Vol 224. 105497. <https://doi.org/10.1016/j.aquatox.2020.105497>.
49. Effects of high hydrostatic pressure and storage temperature on fatty acids and non-volatile taste active compounds in red claw crayfish (*Cherax quadricarinatus*) / Liu C. et al. // *Molecules*. 2022. Vol. 27(16). 5098. <https://doi.org/10.3390/molecules27165098>.
50. Effects of light color, photoperiod, and growth-related gene interference or overexpression on the survival, growth, or physiological and biochemical indices of red claw crayfish juveniles / Cheng S. et al. // *Aquaculture*. 2023. Vol. 562. 738740. <https://doi.org/10.1016/j.aquaculture.2022.738740>
51. Effects of nitrite on physiological indices and intestinal flora of parent *Cherax quadricarinatus* during overwintering / Wei Y. et al. // *Journal of Shanghai Ocean University*. 2022. Vol. 31(1). P. 278—287. <https://doi.org/10.12024/jsou.20201203256>.
52. Energy reserves mobilization: strategies



- chain reaction. *Journal of virological methods*, 293, 114169. <https://doi.org/10.1016/j.jviromet.2021.114169>.
51. Jerry, E. M., Karnaneedi, S., Ruethers, T., Jerry, D. R., Condon, K., & Lopata, A. L. (2024). Allergen diversity and abundance in different tissues of the redclaw crayfish (*Cherax quadricarinatus*). *Foods*, 13(2), 315. <https://doi.org/10.3390/foods13020315>.
52. Jin, L., Jia, S., Zhang, W., Chen, Y., Li, S., Liu, P., Li, J., & Lv, J. (2022). Identification of sex-specific DNA markers: providing molecular evidence for the ZW sex determination system in the red-claw crayfish (*Cherax quadricarinatus*). *Aquaculture*, 546, 737254. <https://doi.org/10.1016/j.aquaculture.2021.737254>.
53. Jutagate, T., Kwangkhwang, W., & Saowakoon, S. (2023). Growth and competitions of the australian red-claw crayfish, *Cherax quadricarinatus* (von Martens, 1868) in Thailand: the experimental approaches. *Aquatic Invasions*, 18(1), 103–117. <https://doi.org/10.3391/ai.2023.18.1.103301>.
54. Kawai, T. (2017). Observation on mandible and gill morphology in *Pacifastacus leniusculus* and *Cherax quadricarinatus* with a review of the introduction of alien crayfish into Japan. *Freshwater Crayfish*, 23, 29–39. <https://doi.org/10.5869/fc.2017.v23-1.29>.
55. King, G., Balcombe, S., Capon, S., & Cockayne, B. (2022). Do opposites attack? Resource interactions between an alien and native crayfish from the lake Eyre basin. *Marine and Freshwater Research*, 73(7), 873–883. <https://doi.org/10.1071/MF21302>.
56. Kodiran, T., Mashar, A., Febriana, R., Nurulhayati, E. R., Nurulhafidzah, A., & Wardiatno, Y. (2020). Development of economic valuation method for the direct impact of alien invasive species based on food competition in aquatic ecosystems. *Jurnal Pengelolaan Sumberdaya Alam Dan Lingkungan*, 10(2), 198–208. <https://doi.org/10.1016/j.jviromet.2017.114169>.
57. Establishing a gold standard method for the detection of *Cherax reovirus* using reverse transcriptase, quantitative, polymerase chain reaction / Jaroenram W. et al. // *Journal of virological methods*. 2021. Vol. 293. 114169. <https://doi.org/10.1016/j.jviromet.2021.114169>.
58. Establishment of a primary culture of haemocytes from the Australian red claw crayfish, *Cherax quadricarinatus* / Li J. et al. // *Crustaceana*. 2019. Vol. 92(11–12). P. 1271–1278. <https://doi.org/10.1163/15685403-00003937>.
59. Establishment of the exotic invasive red-claw crayfish *Cherax quadricarinatus* (Von Martens, 1868) in the coastal plain of San Blas, Nayarit, Se Gulf of California, Mexico / Tapia-Varela J. R. et al. // *BioInvasions Records*. 2020. Vol. 9(2). P. 357–366. <https://doi.org/10.3391/bir.2020.9.2.21>.
60. Estimates of heritability and genetic correlation for growth traits at harvest in redclaw crayfish, *Cherax quadricarinatus* / Dai P. et al. // *Aquaculture*. 2022. Vol. 561. 738631. <https://doi.org/10.1016/j.aquaculture.2022.738631>.
61. Exploration of the functional properties of red cuticle patch of redclaw crayfish (*Cherax quadricarinatus*) by histological and transcriptomic analysis / Zhou S. et al. // *Aquaculture*. 2022. Vol. 561(15). 738624. <https://doi.org/10.1016/j.aquaculture.2022.738624>.
62. Expression patterns of two carbonic anhydrase genes, Na⁺/K⁺-ATPase and V-type H⁺-ATPase, in the freshwater crayfish, *Cherax quadricarinatus*, exposed to low pH and high pH / Ali M. J. et al. // *PLoS One*. 2017. Vol. 12(9). e0184060. <https://doi.org/10.1371/journal.pone.0184060>.



- doi.org/10.29244/jpsl.10.2.198-208.
57. Kruppke, B., Farack, J., Weil, S., Aflalo, E.D., Poláková, D., Sagi, A., & Hanke, T. (2020). Crayfish hemocyanin on chitin bone substitute scaffolds promotes the proliferation and osteogenic differentiation of human mesenchymal stem cells. *Journal of Biomedical Materials Research – Part A*, 108(3), 694–708. <https://doi.org/10.1002/jbm.a.36849>.
 58. Kuhu, R., Mantiri, R. O. S. E., & Tombokan, J. L. (2018). Some biological aspects of freshwater lobsters, *Cherax quadricarinatus*, in Ralik River of Southeast Minahasa and in Tondano Lake of Minahasa. *Jurnal Ilmiah Platax*, 7(1), 34. <https://doi.org/10.35800/jip.7.1.2019.21444>.
 59. Levy, T., & Sagi, A. (2020). The “IAG-Switch” – a key controlling element in *Decapod* crustacean sex differentiation. *Frontiers in Endocrinology*, 11, 651. <https://doi.org/10.3389/fendo.2020.00651>.
 60. Levy, T., Ventura, T., De Leo, G., Grinshpan, N., Abu Abayed, F. A., Manor, R., Savaya, A., Sklarz, M. Y., Chalifa-Caspi, V., Mishmar, D., & Sagi, A. (2020). Two homogametic genotypes – one crayfish: on the consequences of intersexuality. *iScience*, 23, 101652. <https://doi.org/10.1016/j.isci.2020.101652>.
 61. Li, J., Chen, Y., Zhang, T., Jiang, Q., Peng, G., Yang, J., Tang J., & Si, Q. (2019). Establishment of a primary culture of haemocytes from the Australian red claw crayfish, *Cherax quadricarinatus*. *Crustaceana*, 92(11–12), 1271–1278. <https://doi.org/10.1163/15685403-00003937>.
 62. Lin, D., Guo, Y., Chen, X., Yang, H., Li, Q., Liu, Q., Luo, F., Meng, K., Yang, S., Cheng, X., Ma, W., Chen, X., Wang, M., & Zhao, Y. (2022). Identification and expression pattern of the sex determination gene fruitless-like in *Cherax quadricarinatus*. *Comparative Biochemistry and Physiology Part – B: Biochemistry and Molecular Biology*, 259, 110704. <https://doi.org/10.1016/j.cbpb.2021.110704>.
 - Y. et al. // Australian Journal of Zoology. 2017. Vol. 65(1). P. 50—59. <https://doi.org/10.1071/ZO16048>.
 59. Fahrudin M., Suriyadin A., Murtawan H. Pertumbuhan dan kelangsungan hidup lobster air tawar (*Cherax quadricarinatus*) dengan pemberian substrat yang berbeda // Jurnal Marikultur. 2022. Vol. 4(1). P. 31—41.
 60. Fasya A. H. Study of patterns in the relationship of ecdysis with the age of freshwater crayfish *Cherax quadricarinatus* aged 76 days // IOP Conference Series: Earth and Environmental Science. 2019. 236(1). 012012. <https://doi.org/10.1088/1755-1315/236/1/012012>.
 61. Fauzan Isma M., Syahril M. Effects of difference shelter on survival rate and growth of freshwater lobster (*Cherax quadricarinatus*) // Jurnal Ilmiah Samudra Akuatika. 2021. Vol. 5(1). P. 1—8. <https://doi.org/10.33059/jisa.v5i1.3547>.
 62. First record of a new epibionts suctorian ciliate *Tokophrya huangmeiensis* sp.n. (Ciliophora, Phyllopharyngea) from redclaw crayfish *Cherax quadricarinatus* von Martens 1868 / Tahir U. B. et al. // Zootaxa. 2017. Vol. 4269(2). P. 287—295. <https://doi.org/10.11646/zootaxa.4269.2.7>.
 63. First record of the invasive Australian redclaw crayfish *Cherax quadricarinatus* (Von Martens, 1868) in the Crocodile river, Kruger National Park, South Africa / Petersen R. M. et al. // Koedoe: African Protected Area Conservation and Science. 2017. Vol. 59(159). P. 1—3. <https://doi.org/10.4102/koedoe.v59i1.1435>.
 64. First wild record of australian redclaw crayfish *Cherax quadricarinatus* (Von Martens, 1868) in the east coast of peninsular Malaysia / Ismail N. eta al. // Bi-



63. Lin, X., Liu, C., Cai, L., Yang, J., Zhou, J., Jiang, H., Shi, Y., & Gu, Z. (2021). Effect of high hydrostatic pressure processing on biochemical characteristics, bacterial counts, and color of the red claw crayfish *Cherax quadricarinatus*. *Journal of Shellfish Research*, 40, 177–184. <https://doi.org/10.2983/035.040.0117>.
64. Liu, C., Li, M., Wang, Y., Yang, Y., Wang, A., & Gu, Z. (2022). Effects of high hydrostatic pressure and storage temperature on fatty acids and non-volatile taste active compounds in red claw crayfish (*Cherax quadricarinatus*). *Molecules*, 27(16), 5098. <https://doi.org/10.3390/molecules27165098>.
65. Low, B. W., Zeng, Y., Tan, H. H., & Yeo, D. C. J. (2021). Predictor complexity and feature selection affect Maxent model transferability: evidence from global freshwater invasive species. *Diversity and Distributions*, 27(3), 497–511. <https://doi.org/10.1111/ddi.13211>.
66. Lu Y-P., Zheng P-H., Zhang Z-L., Zhang X-X., Li J-T., Wang D-M., Xu J-R., Xian J-A., & Wang A-L. (2023). Hepatopancreas transcriptome alterations in red claw crayfish (*Cherax quadricarinatus*) under microcystin-LR (MC-LR) stress. *Aquaculture Reports*, 29, 101478. <https://doi.org/10.1016/j.aqrep.2023.101478>.
67. Lu, Y. P., Zheng, P. H., Zhang, X. X., Li, J. T., Zhang, Z. L., Xu, J. R., Meng Y-Q., Li, J. J., Xian J. A., & Wang, A. L. (2023). New insights into the regulation mechanism of red claw crayfish (*Cherax quadricarinatus*) hepatopancreas under air exposure using transcriptome analysis. *Fish & Shellfish Immunology*, 132, 108505. <https://doi.org/10.1016/j.fsi.2022.108505>.
68. Macias, N. A., Torres, P. J., & Colón-Gaud, C. (2021). Records of the Australian red-claw crayfish *Cherax quadricarinatus* (von Martens, 1868) on the island of Puerto Rico. *BioInvasions Records*, 10(2), 348–359. <https://doi.org/10.3391/bir.2021.10.2.13>.
69. Madzivanzira, T. C., Chakandinakira, A. oInvasions Records. 2021. Vol. 10(2). P. 360–368. <https://doi.org/10.3391/bir.2021.10.2.14>.
65. First wild record of Australian redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) in the East Coast of Peninsular Malaysia / Norshida I. et al. // BioInvasions Records. 2021. Vol. 10(2). P. 360–368. <https://doi.org/10.3391/bir.2021.10.2.14>.
66. From sporadic single genes to a broader transcriptomic approach: insights into the formation of the biomineralized exoskeleton in decapod crustaceans / Shaked S. A. et al. // Journal of Structural Biology. 2020. Vol. 212(2). 107612. <https://doi.org/10.1016/j.jsb.2020.107612>.
67. Fu R., Li F., Yang F. Separation of hemocytes of *Cherax quadricarinatus* by percoll discontinuous density gradient centrifugation // Journal of Fisheries of China. 2019. Vol. 43(4). P. 841–851. <https://doi.org/10.11964/jfc.20180511286>.
68. Genes encoding putative bicarbonate transporters as a missing molecular link between molt and mineralization in crustaceans / Abehsera S. et al. // Scientific Reports. 2021. Vol. 11(1). 11722. <https://doi.org/10.1038/s41598-021-91155-w>.
69. Genetic variability of wild populations of invasive redclaw crayfish (*Cherax quadricarinatus*) von Martens, 1868 across peninsular Malaysia / Dali M. Z. M. et al. // Malaysian Applied Biology. 2023. Vol. 52(1). P. 35–42. <https://doi.org/10.55230/mabjournal.v52i1.2427>.
70. Genomic structure, expression, and functional characterization of the Fem-1 gene family in the redclaw crayfish, *Cherax quadricarinatus* / Zheng J. et al. // General and comparative endocrinology. 2022. Vol. 316. 113961. <https://doi.org/10.1016/j.ygcen.2022.113961>.



- T., Mungenge, C. P., O'Brien, G., Dalu, T., & South, J. (2023). Get it before it gets to my catch: misdirection traps to mitigate against socioeconomic impacts associated with crayfish invasion. *Management of Biological Invasions*, 14(2), 335–346. <https://doi.org/10.3391/mbi.2023.14.2.10>.
70. Madzivanzira, T. C., South, J., & Weyl, O. L. F. (2021). Invasive crayfish outperform *Potamonautid crabs* at higher temperatures. *Freshwater Biology*, 66(5), 978–991. <https://doi.org/10.1111/fwb.13691>.
71. Madzivanzira, T. C., South, J., Nhwatiwa, T., & Weyl, O. L. F. (2021). Standardisation of alien invasive Australian redclaw crayfish *Cherax quadricarinatus* sampling gear in Africa. *Water SA*, 47(3), 380–384. <https://doi.org/10.17159/wsa/2021.v47.i3.11866>.
72. Madzivanzira, T. C., South, J., Wood, L. E., Nunes, A. L., & Weyl, O. L. F. (2020). A review of freshwater crayfish introductions in Africa. *Reviews in Fisheries Science & Aquaculture*, 29(2), 218–241. <https://doi.org/10.1080/23308249.2020.1802405>.
73. Madzivanzira, T. C., Weyl, O. L. F., & South, J. (2022). Ecological and potential socioeconomic impacts of two globally-invasive crayfish. *NeoBiota*, 72, 25–43. <https://doi.org/10.3897/neobiota.72.71868>.
74. Mamuaya, J., Mingkid, W. M., Kalesaran, O. J., Sinjal, H. J., Tombol, R. A., & Tombokan, J. L. (2019). The survival rate and growth of juvenile crayfish (*Cherax quadricarinatus*) with different types of shelter. *Jurnal Ilmiah Platax*, 7(2), 427–431. <http://dx.doi.org/10.35800/jip.7.2.2019.24510>.
75. Mamuko, N., Mingkid, W. M., Watung, J. C., Kreckhoff, R. L., Longdong, S. N. J., & Manginsela, F. B. (2021). The use of nutmeg oil (*Myristica fragrans*) Houlttuyn 1774 as anesthetic component in different concentrations for the young freshwater lobster (*Cherax quadricarinatus*) Von Martens 1868. *E-Journal Budidaya Perairan*, 10(1), 73–80. <https://doi.org/10.1016/j.ygcen.2021.113961>.
71. Get it before it gets to my catch: misdirection traps to mitigate against socioeconomic impacts associated with crayfish invasion / Madzivanzira T. C. et al. // Management of Biological Invasions. 2023. Vol. 14(2). P. 335–346. <https://doi.org/10.3391/mbi.2023.14.2.10>.
72. Haemolymphatic parameters in two aquaculture crustacean species *Cherax destructor* (Clark, 1836) and *Cherax quadricarinatus* (Von Martens, 1868) / Mauro M. et al. // Animals. 2022. Vol. 12(5). P. 543. <https://doi.org/10.3390/ani12050543>.
73. Hepatopancreas transcriptome alterations in red claw crayfish (*Cherax quadricarinatus*) under microcystin-LR (MC-LR) stress / Lu Y-P. et al. // Aquaculture Reports. 2023. Vol. 29. 101478. <https://doi.org/10.1016/j.aqrep.2023.101478>.
74. Identification and expression pattern of the sex determination gene fruitless-like in *Cherax quadricarinatus* / Lin D. et al. // Comparative Biochemistry and Physiology Part — B: Biochemistry and Molecular Biology. 2022. Vol. 259. 110704. <https://doi.org/10.1016/j.cbpb.2021.110704>.
75. Identification and functional analysis of the doublesex gene in the redclaw crayfish, *Cherax quadricarinatus* / Zheng J. et al. // Gene expression patterns. 2020. Vol. 37. 119129. <https://doi.org/10.1016/j.gexp.2020.119129>.
76. Identification and profiling of MicroRNAs during embryogenesis in the red claw crayfish *Cherax quadricarinatus* / Wang Y. et al. // Frontiers in physiology. 2020. Vol. 11. P. 878. <https://doi.org/10.3389/fphys.2020.00878>.
77. Identification of sex-specific DNA



- doi.org/10.35800/bdp.10.1.2022.35535.
76. Marshall, B. E. (2019). Crayfish, catfish and snails: the perils of uncontrolled biological control. *African Journal of Aquatic Science*, 44(1), 1–5. <https://doi.org/10.2989/16085914.2019.1599810>.
77. Marufu, L. T., Dalu, T., Crispen, P., Barson, M., Simango, R., Utete, B., & Nhiwatiwa, T. (2018). The diet of an invasive crayfish, *Cherax quadricarinatus* (Von Martens, 1868), in Lake Kariba, inferred using stomach content and stable isotope analyses. *BioInvasions Records*, 7(2), 121–132. <https://doi.org/10.3391/bir.2018.7.2.03>.
78. Marufu, L., Dalu, T., Phiri, C., & Nhiwatiwa, T. (2017). Diet composition changes in tigerfish of Lake Kariba following an invasion by redclaw crayfish. *Annales de Limnologie*, 53, 47–56. <https://doi.org/10.1051/limn/2016033>.
79. Mashar, A., Wahyuni, Y. S., Hakim, A. A., & Wardiatno, Y. (2019). Pendekatan truss morfometric dalam menganalisis kekerabatan populasi *Cherax quadricarinatus* (Von Martens, 1868) di Perairan Jawa Barat. *Jurnal Pengelolaan Perikanan Tropis*, 3(2), 20–27. <http://dx.doi.org/10.29244/jppt.v3i2.30432>.
80. Mauro, M., Arizza, V., Arculeo, M., Attanzio, A., Pinto, P., Chirco, P., Badalamenti G., Tesoriere L., & Vazzana, M. (2022). Haemolymphatic parameters in two aquaculture crustacean species *Cherax destructor* (Clark, 1836) and *Cherax quadricarinatus* (Von Martens, 1868). *Animals*, 12(5), 543. <https://doi.org/10.3390/ani12050543>.
81. Meng, F., Zhang, G., Yin, W., Wang, L., & Sun, J. (2019). Study on interaction of autophagosome and microtubule during autophagy process in *Cherax quadricarinatus*. *Journal of Fisheries of China*, 43(12), 2494–2500. <https://doi.org/10.11964/jfc.20180811406>.
82. Mhlanga, L., Marufu, L., Mupandawana, G., & Nhiwatiwa, T. (2020). An examination of the effectiveness of traps and baits as a possible means of harvest markers: providing molecular evidence for the ZW sex determination system in the redclaw crayfish (*Cherax quadricarinatus*) / Jin L. et al. // *Aquaculture*. 2022. Vol. 546. 737254. <https://doi.org/10.1016/j.aquaculture.2021.737254>.
78. In a pinch: mechanisms behind potential biotic resistance toward two invasive crayfish by native African freshwater crabs / South J. et al. // *Frontiers in Ecology and Evolution*. 2020. Vol. 8. P. 72. <https://doi.org/10.3389/fevo.2020.00072>.
79. Influence of salinity on survival, growth, hemolymph osmolality, gill sodium potassium ATPase activity, and sodium potassium chloride co-transporter expression in the redclaw crayfish *Cherax quadricarinatus* / Rida R. et al. // *Journal of the World Aquaculture Society*. 2021. Vol. 52(2). P. 466–474. <https://doi.org/10.1111/jwas.12762>.
80. Invasion and distribution of the redclaw crayfish, *Cherax quadricarinatus*, in Martinique / Baudry T. et al. // *Knowledge and Management of Aquatic Ecosystems*. 2020. Vol. 421. P. 50. <https://doi.org/10.1051/kmae/2020041>.
81. Jutagate T., Kwangkhwang W., Saowakoon S. Growth and competitions of the Australian red-claw crayfish, *Cherax quadricarinatus* (von Martens, 1868) in Thailand: the experimental approaches // *Aquatic Invasions*. 2023. Vol. 18(1). P. 103–117. <https://doi.org/10.3391/ai.2023.18.1.103301>.
82. Kawai T. Observation on mandible and gill morphology in *Pacifastacus leniusculus* and *Cherax quadricarinatus* with a review of the introduction of alien crayfish into Japan // *Freshwater Crayfish*. 2017. Vol. 23. P. 29–39. <https://doi.org/10.5869/fc.2017.v23-1.29>.



- ing crayfish, *Cherax quadricarinatus* in Sanyati Basin, Lake Kariba, Zimbabwe. *Water SA*, 46(4), 675–678. <https://doi.org/10.17159/wsa/2020.v46.i4.9083>.
83. Minh Nhut, T., Mykles, D.L., Elizur, A., & Ventura, T. (2020). Ecdysis triggering hormone modulates molt behaviour in the redclaw crayfish *Cherax quadricarinatus*, providing a mechanistic evidence for conserved function in molt regulation across *Pan crustacea*. *General and Comparative Endocrinology*, 298(1), 113556. <https://doi.org/10.1016/j.ygcen.2020.113556>.
84. Mornigstar, C. R., Daniel, W. M., Neilson, M. E., & Yazaryan, A. K. (2020). The first occurrence of the Australian redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) in the contiguous United States. *BioInvasions Records*, 9(1), 120–126. <https://doi.org/10.3391/bir.2020.9.1.16>.
85. Nasir, S. A. M., Khaleel, A. G., Badaluddin, N. A., Kok, S.-Y., Ismail, N., Sheriff, S. M., & Kamarudin, A.-S. (2020). Development of species-specific primer sets for Australian redclaw crayfish (*Cherax quadricarinatus*) detection from water environmental DNA (eDNA). *Bioscience Research*, 17, 90–99.
86. Neelima, H., Srinivasa, R. B., & Ramachandra, R. P. (2017). Crustacean molting: regulation and effects of environmental toxicants. *Journal of Marine Science: Research & Development*, 7(5), 236. <http://dx.doi.org/10.4172/2155-9910.1000236>.
87. Nguyen, C. D. H., Ventura, T., & Elizur, A. (2019). Crustacean nuclear localization signals help facilitating the delivery of DNA into Australian red-claw crayfish cells. *Aquaculture*, 499, 149–159. <https://doi.org/10.1016/j.aquaculture.2018.09.030>.
88. Nong, C., Chen, Y., Yang, H., Chen, N., Tian, C., Li, S., & Chen, H. (2024). Phenotypic sorting of individual male and female intersex *Cherax quadricarinatus* and analysis of molecular differences in the gonadal transcriptome. *Comparative Biochem-*
83. Kuhu R., Mantiri R. O. S. E., Tombokan J. L. Some biological aspects of freshwater lobsters, *Cherax quadricarinatus*, in Ralik River of Southeast Minahasa and in Tondano Lake of Minahasa // *Jurnal Ilmiah Platax*. 2018. Vol. 7(1). P. 34. <https://doi.org/10.35800/jip.7.1.2019.21444>.
84. Levy T., Sagi A. The “IAG-Switch” — a key controlling element in Decapod crustacean sex differentiation // *Frontiers in Endocrinology*. 2020. Vol. 11. P. 651. <https://doi.org/10.3389/fendo.2020.00651>.
85. Macias N. A., Torres P. J., Colón-Gaud C. Records of the Australian redclaw crayfish *Cherax quadricarinatus* (Von Martens, 1868) on the island of Puerto Rico // *BioInvasions Records*. 2021. Vol. 10(2). P. 348–359. <https://doi.org/10.3391/bir.2021.10.2.13>.
86. Madzivanzira T. C., South J., Weyl O. L. F. Invasive crayfish outperform *Potamonautid crabs* at higher temperatures // *Freshwater Biology*. 2021. Vol. 66(5). P. 978–991. <https://doi.org/10.1111/fwb.13691>.
87. Madzivanzira T. C., Weyl O. L. F., South J. Ecological and potential socio-economic impacts of two globally-invasive crayfish // *NeoBiota*. 2022. Vol. 72. P. 25–43. <https://doi.org/10.3897/neobiota.72.71868>.
88. Marshall B. E. Crayfish, catfish and snails: the perils of uncontrolled biological control // *African Journal of Aquatic Science*. 2019. Vol. 44(1). P. 1–5. <https://doi.org/10.2989/16085914.2019.1599810>.
89. Mobilization of energetic reserves during starvation in juveniles of different size of the redclaw crayfish *Cherax quadricarinatus* / Calvo N. S. et al. //



- istry and Physiology – Part D: Genomics and Proteomics*, 49, 101194. <https://doi.org/10.1016/j.cbd.2024.101194>.
89. Nong, W., Chai, Z. Y. H., Jiang, X., Qin, J., Ma, K. Y., Chan, K. M., Chan, T. F., Chow, B. K. C., Kwan, H. S., Wong, C. K. C., Qiu, J. W., Hui, J. H. L., & Chu, K. H. (2020). A crustacean annotated transcriptome (CAT) database. *BMC Genomics*, 21(1), 32. <https://doi.org/10.1186/s12864-019-6433-3>.
90. Norshida, I., Mohd Nasir, M. S. A., Khaleel, A. G., Sallehuddin, A. S., Syed Idrus, S. N., Istiqomah, I., & Ahmad, S. K. (2021). First wild record of Australian redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) in the East Coast of Peninsular Malaysia. *BioInvasions Records*, 10(2), 360–368. <https://doi.org/10.3391/bir.2021.10.2.14>.
91. Nur, M., Komariyah, S., & Haser, T. F. (2023). The influence of various substrates on the survival of freshwater lobster (*Cherax quadricarinatus*) in the open transport system. *Jurnal Ilmiah Samudra Akuatika*, 7(2), 1–8. <https://doi.org/10.33059/jisa.v7i2.9022>.
92. Patoka, J., Wardiatno, Y., Mashar, A., Wowor, D., Jerikho, R., Takdir, M., Purnamasari, L., Petryl, M., Kalous, L., Kouba, A., & Blaha, M. (2018). Redclaw crayfish, *Cherax quadricarinatus* (von Martens, 1868), widespread throughout Indonesia. *BioInvasions Records*, 7(2), 185–189. <https://doi.org/10.3391/bir.2018.7.2.11>.
93. Petersen, R. M., Hoffman, A. C., Kotze, P., & Marr, S. M. (2017). First record of the invasive Australian redclaw crayfish *Cherax quadricarinatus* (Von Martens, 1868) in the Crocodile river, Kruger National Park, South Africa. *Koedoe: African Protected Area Conservation and Science*, 59(159), 1–3. <https://doi.org/10.4102/koedoe.v59i1.1435>.
94. Purnamasari, L., Kasmeri, R., Amin, M. H. F. adil, & Soegianto, A. (2018). Aquaculture Nutrition. 2018. Vol. 24(3). P. 952—960. <https://doi.org/10.1111/anu.12631>.
90. Modeling the habitat suitability of two exotic freshwater crayfishes in Mesoamerica and the Caribbean: *Cherax quadricarinatus* (von Martens, 1868) and *Procambarus clarkii* Girard, 1852 (Decapoda: Astacidea: Parastacidae, Cambaridae) / Azofeifa-Solano J. C. et al. // Journal of Crustacean Biology. 2023. Vol. 43(4). ruad059. <https://doi.org/10.1093/jcobiol/ruad059>.
91. Molecular characterization and expression profiling of three transformer-2 splice isoforms in the redclaw crayfish, *Cherax quadricarinatus* / Cai L. et al. // Frontiers in Physiology. 2020. Vol. 11(631). 10. <https://doi.org/10.3389/fphys.2020.00631>.
92. Molecular characterization of the Ftz-fl gene in redclaw crayfish *Cherax quadricarinatus* and its potential role in ovarian development / Chen L. et al. // Aquaculture Research. 2022. Vol. 53(15). P. 5261—5269. <https://doi.org/10.1111/are.16010>.
93. Molecular identification and expression profiles of four splice variants of sex-lethal gene in *Cherax quadricarinatus* / Zheng J. et al. // Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology. 2019. Vol. 234. P. 26—33. <https://doi.org/10.1016/j.cbpb.2019.05.002>.
94. Morphological and molecular identification of *Diceratocephala boschmai* Baer, 1953 and *Decadidymus* sp. Cannon, 1991 on wild and cultured environment of *Cherax quadricarinatus* in Malaysia / Hassan M. et al. // BioInvasions Records. 2022. Vol. 11(2). P. 495—509. <https://doi.org/10.3391/>



- Morphometric characteristics of alien crayfish *Cherax quadricarinatus* from Maninjau Lake (West Sumatra, Indonesia). *Polish Journal of Natural Sciences*, 33(3), 369–383.
95. Rakhmawati, R., Aprilia, T., Indariyanti, N., Saputra, A., & Marlina, E. (2023). Color quality improvement of *Cherax quadricarinatus* with dragon fruit peel meal utilization. *Depik*, 12(3), 314–319. <https://doi.org/10.13170/depik.12.3.28628>.
96. Rani, Z., Ridwanto, Nasution, H. M., Kaban, V. E., Nasri, N., & Karo, N. B. (2023). Antibacterial activity of freshwater lobster (*Cherax quadricarinatus*) shell chitosan gel preparation against *Escherichia coli* and *Staphylococcus aureus*. *Journal of Applied Pharmaceutical Science*, 13(2), 146–153. <https://doi.org/10.7324/JAPS.2023.130216>.
97. Rida, R., Zein-Eddine, R., Kreydiyyeh, S., Garza de Yta, A., & Saoud, I. P. (2021). Influence of salinity on survival, growth, hemolymph osmolality, gill sodium potassium ATPase activity, and sodium potassium chloride co-transporter expression in the redclaw crayfish *Cherax quadricarinatus*. *Journal of the World Aquaculture Society*, 52(2), 466–474. <https://doi.org/10.1111/jwas.12762>.
98. Ridwanto, Daulay, A. S., & Gurning, K. (2022). Isolation and characterization of chitosan from sea and freshwater waste, North Sumatera province, Indonesia. *Rasayan Journal of Chemistry*, 15(2), 780–785. <https://doi.org/10.31788/RJC.2022.1526721>.
99. Rigg, D. P., Courtney, R. L., Jones, C. M., & Seymour, J. E. (2021). Morphology and weight-length relationships for the first six instars of *Cherax quadricarinatus* (von Martens, 1868). *Freshwater Crayfish*, 26(1), 9–16. <https://doi.org/10.5869/fc.2021.v26-1.9>.
100. Rodríguez-Almaraz, G. A., Mendoza, R., Aguilera-González, C., Barriga, C., & Tirado-Velarde, M. (2018). Additional bir.2022.11.2.22.
95. Morphology and weight-length relationships for the first six instars of *Cherax quadricarinatus* (von Martens, 1868) / Rigg D. P. et al. // *Freshwater Crayfish*. 2021. Vol. 26(1). P. 9—16. <https://doi.org/10.5869/fc.2021.v26-1.9>.
96. Morphometric characteristics of alien crayfish *Cherax quadricarinatus* from Maninjau Lake (West Sumatra, Indonesia) / Purnamasari L. et al. // *Polish Journal of Natural Sciences*. 2018. Vol. 33(3). P. 369—383.
97. Neelima H., Srinivasa R. B., Ramachandra R. P. (2017). Crustacean molting: regulation and effects of environmental toxicants // *Journal of Marine Science: Research & Development*. 2017. Vol. 7(5). P. 236. <http://dx.doi.org/10.4172/2155-9910.1000236>.
98. Neuroanatomy and morphological diversity of brain cells from adult crayfish *Cherax quadricarinatus* / Duan H. et al. // *Journal of Oceanology and Limnology*. 2018. Vol. 36(6). P. 2368—2378. <https://doi.org/10.1007/s00343-019-7145-x>.
99. New insights into the regulation mechanism of red claw crayfish (*Cherax quadricarinatus*) hepatopancreas under air exposure using transcriptome analysis / Lu Y. P. et al. // *Fish & Shellfish Immunology*. 2023. Vol. 132. 108505. <https://doi.org/10.1016/j.fsi.2022.108505>.
100. Nguyen C. D. H., Ventura T., Elizur A. Crustacean nuclear localization signals help facilitating the delivery of DNA into Australian red-claw crayfish cells // *Aquaculture*. 2019. Vol. 499. P. 149—159. <https://doi.org/10.1016/j.aquaculture.2018.09.030>.
101. Nur M., Komariyah S., Haser T. F. The



- records of wild populations of the Australian crayfish *Cherax quadricarinatus* in Mexico. *Revista Mexicana de Biodiversidad*, 89(4), 1322–1327. <https://doi.org/10.22201/ib.20078706e.2018.4.2065>.
101. Sacristán, H. J., Rodríguez, Y. E., De Los Angeles Pereira, N., López Greco, L. S., Lovrich, G. A., & Fernández Gimenez, A. V. (2017). Energy reserves mobilization: strategies of three decapod species. *PLoS One*, 12(9), e0184060. <https://doi.org/10.1371/journal.pone.0184060>.
102. Sakuna, K., Elliman, J., & Owens, L. (2018). Comparison of molecular detection PCR methods for *Chequua iflavirus* in freshwater crayfish, *Cherax quadricarinatus*. *Journal of virological methods*, 251, 139–144. <https://doi.org/10.1016/j.jviromet.2017.10.013>.
103. Sánchez-Hernández, J. C., Pacheco-Ortiz, J. A., Rodríguez-Sosa, L., Calderón-Rosete, G., & Villagran-Vargas, E. (2018). Asymmetric firing rate from crayfish left and right caudal photoreceptors due to blue and green monochromatic light pulses. *Symmetry*, 10(9), 389. <https://doi.org/10.3390/sym10090389>.
104. Sánchez-Salgado, J. L., Pereyra, M. A., Agundis, C., Vivanco-Rojas, O., Rosales, C., Pascual, C., Alpuche-Osorno, J. J., & Zenteno, E. (2018). The effect of the lectin from *Cherax quadricarinatus* on its granular hemocytes. *Fish & Shellfish Immunology*, 77, 131–138. <https://doi.org/10.1016/j.fsi.2018.03.050>.
105. Sanjar, A., Davis, D. R., & Kline, R. J. (2023). Evidence of an established population of *Cherax quadricarinatus* (von Martens, 1868) in south Texas, USA. *BioInvasions Records*, 12(1), 284–291. <https://doi.org/10.3391/bir.2023.12.1.24>.
106. Shahroom, A., Shapawi, R., Mustafa, S., Halid, N. F. A., Estim, A., & Tuzan, A. D. (2023). Effects of eggshell and seashell powder as natural dietary calcium supplements on growth, molting frequency, and carapace calcium composition of various substrates on the survival of freshwater lobster (*Cherax quadricarinatus*) in the open transport system // Jurnal Ilmiah Samudra Akuatika. 2023. Vol. 7(2). P. 1–8. <https://doi.org/10.33059/jisa.v7i2.9022>.
102. Nutritional composition and antioxidant activity of freshwater lobster in Malaysia: a short review / Tengku Zainal Abidin T. Z. H. et al. // Asian Journal of Chemistry. 2023. Vol. 35(2). P. 301–304. <https://doi.org/10.14233/ajchem.2023.24050>.
103. Pendekatan truss morfometric dalam menganalisis kekerabatan populasi *Cherax quadricarinatus* (Von Martens, 1868) di Perairan Jawa Barat / Mashar A. et al. // Jurnal Pengelolaan Perikanan Tropis. 2019. Vol. 3(2). P. 20–27. <http://dx.doi.org/10.29244/jppt.v3i2.30432>.
104. Phenotypic sorting of individual male and female intersex *Cherax quadricarinatus* and analysis of molecular differences in the gonadal transcriptome / Nong, C. et al. // Comparative Biochemistry and Physiology — Part D: Genomics and Proteomics. 2024. Vol. 49. 101194. <https://doi.org/10.1016/j.cbd.2024.101194>.
105. Predicting *Cherax quadricarinatus* habitat distribution patterns through the usage of GIS and eDNA analysis in Terengganu, Malaysia / Dali M. Z. M. et al. // Sains Malaysiana. 2023. Vol. 52(2). P. 343–354. <https://doi.org/10.17576/jsm-2023-5202-03>.
106. Predictor complexity and feature selection affect Maxent model transferability: evidence from global freshwater invasive species / Low B. W. et al. // Diversity and Distributions. 2021. Vol. 27(3). P. 497–511. <https://doi.org/10.1111/ddi.13211>.



- tion of juvenile red claw crayfish, *Cherax quadricarinatus*. *PeerJ*, 11, e15449. <https://doi.org/10.7717/peerj.15449>.
107. Shaked, S. A., Abehsera, S., Levy, T., Chalifa-Caspi, V., & Sagi, A. (2020). From sporadic single genes to a broader transcriptomic approach: insights into the formation of the biomineralized exoskeleton in decapod crustaceans. *Journal of Structural Biology*, 212(2), 107612. <https://doi.org/10.1016/j.jsb.2020.107612>.
 108. Shaked, S. A., Abehsera, S., Ziegler, A., Bentov, S., Manor, R., Weil, S., Ohana, E., Eichler, J., Aflalo, E. D., & Sagi, A. (2024). A transporter that allows phosphate ions to control the polymorph of exoskeletal calcium carbonate biomineralization. *Acta Biomaterialia*, 178, 221–232. <https://doi.org/10.1016/j.actbio.2024.02.035>.
 109. Shehata, A. I., Wang, T., Habib, Y. J., Wang, J., Fayed, W. M., & Zhang, Z. (2020). The combined effect of vitamin E, arachidonic acid, *Haemtococcus pluvialis*, nucleotides and yeast extract on growth and ovarian development of crayfish (*Cherax quadricarinatus*) by the orthogonal array design. *Aquaculture Nutrition*, 26(6), 2007–2022. <https://doi.org/10.1111/anu.13142>.
 110. Shehata, A. I., Wang, T., Wang, J., & Zhang, Z. (2022). Effect of feed additives in the diet on the growth and testicular development of male red claw crayfish (*Cherax quadricarinatus*) using orthogonal experiments. *Animal Feed Science and Technology*, 283, 115180. <https://doi.org/10.1016/j.anifeedsci.2021.115180>.
 111. Shyamal, S., Das, S., Guruacharya, A., Mykles, D. L., & Durica, D. S. (2018). Transcriptomic analysis of crustacean molting gland (Y-organ) regulation via the mTOR signaling pathway. *Scientific Reports*, 8, 7307. <https://doi.org/10.1038/s41598-018-25368-x>.
 112. Sidharta, V., Pinandoyo, P., & Nugroho, R. A. (2018). Performa kematangan gonad, fekunditas, dan derajat penetasan
 107. Presence of the Australian redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) (Parastacidae, Astacoidea) in a freshwater system in the Caribbean drainage of Costa Rica / Azofeifa-Solano J. C. et al. // *BioInvasions Records*. 2017. Vol. 6(4). P. 351–355. <https://doi.org/10.3391/bir.2017.6.4.08>.
 108. Prevalence of temnocephalids on cultured and wild *Cherax quadricarinatus* in Malaysia / Hassan M. et al. // *Egyptian Journal of Aquatic Research*. 2023. Vol. 49(3). P. 395–400. <https://dx.doi.org/10.1016/j.ejar.2022.03.004>.
 109. Produksi telur *Cherax quadricarinatus* (Von Martens, 1868) di danau Lido, Jawa Barat / Iqbal M. A. et al. // *Jurnal Pengelolaan Perikanan Tropis*. 2019. Vol. 3(2). P.45–52.
 110. Publisher Correction: The chromosome-level genome of *Cherax quadricarinatus* / Chen H. et al. // *Scientific Data*. 2023. Vol. 10(1). P. 313. <https://doi.org/10.1038/s41597-023-02186-z>.
 111. Red claw crayfish *Cherax quadricarinatus* cultivation influences the dynamics and assembly of benthic bacterial communities in paddy fields / Hou Y. et al. // *Environments*. 2023. Vol. 10(10). P. 178. <https://doi.org/10.3390/environments10100178>.
 112. Redclaw crayfish (*Cherax quadricarinatus*): spatial distribution and dispersal pattern in Java, Indonesia / Akmal S. G. et al. // *Knowledge and Management of Aquatic Ecosystems*. 2021. Vol. 422. P. 16. <https://doi.org/10.1051/kmae/2021015>.
 113. Redclaw crayfish, *Cherax quadricarinatus* (von Martens, 1868), widespread throughout Indonesia / Patoka J. et al. // *BioInvasions Records*. 2018. Vol. 7(2). P. 185–189. <https://doi.org/10.3391/>



- melalui strategi pemberian pakan alami yang berbeda pada calon induk lobster air tawar (*Cherax quadricarinatus*). *Jurnal Sains Akuakultur Tropis*, 2(2), 64–74. <https://doi.org/10.14710/sat.v2i2.3150>.
113. Slusar, M., Muzhenko, A., Kovalchuk, I., Borshchenko, V., & Verbelchuk, T. (2023). Study of the embryonic period of female crayfish egg development in different species. *Scientific Horizons*, 26(12), 22–31. <https://doi.org/10.48077/scihor12.2023.22>.
114. Smith, G., Glendinning, S., & Ventura, T. (2023). Transcriptomic changes following induced de-masculinisation of australian red claw crayfish *Cherax quadricarinatus*. *International Journal of Molecular Sciences*, 24(4), 3292. <https://doi.org/10.3390/ijms24043292>.
115. South, J., Madzivanzira, T. C., Tshali, N., Measey, J., & Weyl, O. L. F. (2020). In a pinch: mechanisms behind potential biotic resistance toward two invasive crayfish by native African freshwater crabs. *Frontiers in Ecology and Evolution*, 8, 72. <https://doi.org/10.3389/fevo.2020.00072>.
116. Sun, L. H., Li, Q., Jiang, J. H., Chen, J. M., Gao, L. M., & Guo, J. L. (2023). Analysis on phenotypic traits and muscle nutritional composition of *Cherax quadricarinatus* in different specifications. *Oceanologia et Limnologia Sinica*, 54(3), 885–894. <https://doi.org/10.11693/hyhz20220800220>.
117. Sun, L., Li, Q., Zhang, H., Jiang, J., Chen, J., Gao, L., & Guo, J. (2023). Analysis of change of nutrients from fertilized eggs to newly hatched shrimp of *Cherax quadricarinatus*. *Journal of Shanghai Ocean University*, 32(2), 348–356. <https://doi.org/10.12024/jsou.20220203718>.
118. Sun, Y., Shan, X., Li, D., Liu, X., Han, Z., Qin, J., Guan B., Tan L., Zheng J., Wei M., & Jia, Y. (2023). Analysis of the differences in muscle nutrition among individuals of different sexes in red-claw crayfish, *Cherax quadricarinatus*. *Metabolites*, 13(2), 190. <https://doi.org/10.3390/met13020190>.
119. Sanjar A., Davis D. R., Kline R. J. Evidence of an established population of *Cherax quadricarinatus* (von Martens, 1868) in south Texas, USA // *BioInvasions Records*. 2023. Vol. 12(1). P. 284–291. <https://doi.org/10.3391/bir.2023.12.1.24>.
120. Shelters presence effect on development of early juvenile *Cherax quadricarinatus* (Decapoda; Parastacidae) / *bir*.2018.7.2.11.
114. Reservoirs act as footholds for an invasive freshwater crayfish / Beatty S. J. et al. // *Pacific Conservation Biology*. 2020. Vol. 26(1). P. 78–83. <https://doi.org/10.1071/PC19012>.
115. Reverse transcription polymerase chain reaction (RT-PCR) detection for Australian *Cherax reovirus* from red-claw crayfish (*Cherax quadricarinatus*) / Hayakijkosol O. et al. // *Aquaculture*. 2021. Vol. 530. 735881. <https://doi.org/10.1016/j.aquaculture.2020.735881>.
116. Ridwanto Daulay A. S., Gurning K. Isolation and characterization of chitosan from sea and freshwater waste, North Sumatera province, Indonesia // *Rasayan Journal of Chemistry*. 2022. Vol. 15(2). P. 780–785. <https://doi.org/10.31788/RJC.2022.1526721>.
117. Risk assessment of pet-traded decapod crustaceans in Hungary with evidence of *Cherax quadricarinatus* (von Martens, 1868) in the wild / Weiperth A. et al. // *North-Western Journal of Zoology*. 2019. Vol. 15(1). P. 42–47.
118. Sakuna K., Elliman J., Owens L. Comparison of molecular detection PCR methods for *Chequa iflavirus* in freshwater crayfish, *Cherax quadricarinatus* // *Journal of virological methods*. 2018. Vol. 251. P. 139–144. <https://doi.org/10.1016/j.jviromet.2017.10.013>.



- org/10.3390/metabo13020190.
119. Susanto, G. N., Supono, & Ikrom, F. D. (2018). Sex reversal of juvenile freshwater crayfish (*Cherax quadricarinatus*) influenced by steroid extract of sea cucumber and 17 α -methyltestosterone hormone at different temperatures. *In AIP Conference Proceedings, 2002(1)*. <https://doi.org/10.1063/1.5050123>.
120. Syaharuddin, S. (2021). Pengaruh penambahan kalsium karbonat (CaCO₃) terhadap kelangsungan hidup benih lobster air tawar (*Cherax quadricarinatus*). *Agrokompleks, 21(2)*, 48–52. <https://doi.org/10.51978/japp.v21i2.369>.
121. Tahir, U. B., Deng, Q., Li, S., Liu, Y., Wang, Z. & Gu, Z. (2017). First record of a new epibionts suctorian ciliate *Tokophrya huangmeiensis* sp.n. (Ciliophora, Phyllopharyngea) from red-claw crayfish *Cherax quadricarinatus* von Martens 1868. *Zootaxa, 4269(2)*, 287–295. <https://doi.org/10.11646/zootaxa.4269.2.7>.
122. Tampubolon, I., & Maitindom, F. A. (2023). Length weight relationship of fresh water lobster (*Cherax quadricarinatus*) in lake Paniai, Paniai district. *Jurnal Cakrawala Ilmiah, 2(8)*, 3251–3260. <https://doi.org/10.53625/jcijurnal-cakrawalailmiah.v2i8.5518>.
123. Tan, M. H., Gan, H. M., Lee, Y. P., Grandjean, F., Croft, L. J., & Austin, C. M. (2020). A giant genome for a giant crayfish (*Cherax quadricarinatus*) with insights into cox1 pseudogenes in decapod genomes. *Frontiers in genetics, 11*, 201. <https://doi.org/10.3389/fgene.2020.00201>.
124. Tapia-Varela, J. R., Ponce-Palafox, J. T., Palacios-Salgado, D. S., Romero-Bañuelos, C. A., Nieto-Navarro, J. T., & Aguiar-García, P. (2020). Establishment of the exotic invasive redclaw crayfish *Cherax quadricarinatus* (Von martens, 1868) in the coastal plain of San Blas, Nayarit, Se Gulf of California. *Hernández-Rubio M. C. et al. // Hidrobiologica. 2021. Vol. 30(3). P. 69–75. <https://doi.org/10.24275/uam/izt/dcbs/hidro/2021v31n1/Hernandez>*.
121. Sidharta V., Pinandoyo P., Nugroho R. A. Performa kematangan gonad, fekunditas, dan derajat penetasan melalui strategi pemberian pakan alami yang berbeda pada calon induk lobster air tawar (*Cherax quadricarinatus*) // *Jurnal Sains Akuakultur Tropis. 2018. Vol. 2(2). P. 64–74. <https://doi.org/10.14710/sat.v2i2.3150>*.
122. Smith G., Glendinning S., Ventura T. Transcriptomic changes following induced de-masculinisation of Australian red claw crayfish *Cherax quadricarinatus* // *International Journal of Molecular Sciences. 2023. Vol. 24(4). P. 3292. <https://doi.org/10.3390/ijms24043292>*.
123. Socioeconomic impacts of Australian redclaw crayfish *Cherax quadricarinatus* in Lake Kariba / Chakandinakira A. T. et al. // *Biological Invasions. 2023. Vol. 25(9). P. 2801–2812. <https://doi.org/10.1007/s10530-023-03074-8>*.
124. Standardisation of alien invasive Australian redclaw crayfish *Cherax quadricarinatus* sampling gear in Africa / Madzivanzira T. C. et al. // *Water SA. 2021. Vol. 47(3). P. 380–384. <https://doi.org/10.17159/wsa/2021.v47.i3.11866>*.
125. Study of the embryonic period of female crayfish egg development in different species / Slusar M. et al. // *Scientific Horizons. 2023. Vol. 26(12). P. 22–31. <https://doi.org/10.48077/scihor12.2023.22>*.
126. Study on interaction of autophagosome and microtubule during autophagy process in *Cherax quadricarinatus* / Meng F. et al. // *Journal of Fish-*



- nia, Mexico. *BioInvasions Records*, 9(2), 357–366. <https://doi.org/10.3391/bir.2020.9.2.21>.
125. Taryono, Mashar, A., & Aryasa, S. (2021). Management policy of invasive species red claw crayfish (*Cherax quadricarinatus*) at Lido Lake, Bogor Regency. In *IOP Conference Series: Earth and Environmental Science*, 744, 012090. <https://doi.org/10.1088/1755-1315/744/1/012090>.
126. Tee, Z-B., Ibrahim, S., & Teoh, C-Y. (2022). Comparative study on the nutritional content and physical attributes of giant freshwater prawn (*Macrobrachium rosenbergii*) and redclaw crayfish (*Cherax quadricarinatus*) meats. 07 June. Preprint (Version 1) available at *Research Square*. <https://doi.org/10.21203/rs.3.rs-1695209/v1>.
127. Tengku Zainal Abidin, T. Z. H., Mat Ali, N. N., Abu Bakar, F. I., Ahmad Bakri, F. A., Abu Bakar, M. F., Malik, N. H., & Abidin, M. Z. (2023). Nutritional composition and antioxidant activity of freshwater lobster in Malaysia: a short review. *Asian Journal of Chemistry*, 35(2), 301–304. <https://doi.org/10.14233/ajchem.2023.24050>.
128. Tierney, L. J., Wild, C. H., & Furse, J. M. (2019). Total incombustible (mineral) content of *Cherax quadricarinatus* differs between feral populations in Central-Eastern Australia. *PeerJ*, 17. <http://dx.doi.org/10.7717/peerj.6351>.
129. Tiftazani, M. H., Suparmi, & Sumarto. (2022). Storage time of freshwater lobster (*Cherax quadricarinatus*) with green betel leaf anesthesia (*Piper betle L.*). *Terubuk*, 50(3), 1619–1623.
130. Todd, S.-R.L., & Hyslop, E. J. (2023). Morphometrics, diet, reproductive biology, and ecological interactions of the introduced redclaw crayfish, *Cherax quadricarinatus* (Decapoda: Parastacidae), in Jamaica, West Indies. *Caribbean Journal of Science*, 53(2), 397–410.
- eries of China. 2019. Vol. 43(12). P. 2494—2500. <https://doi.org/10.11964/jfc.20180811406>.
127. Susanto G. N., Supono Ikrom F. D. Sex reversal of juvenile freshwater crayfish (*Cherax quadricarinatus*) influenced by steroid extract of sea cucumber and 17 α -methyltestosterone hormone at different temperatures // In *AIP Conference Proceedings*. 2018. 2002(1). <https://doi.org/10.1063/1.5050123>.
128. Syaharuddin S. Pengaruh penambahan kalsium karbonat (CaCO₃) terhadap kelangsungan hidup benih lobster air tawar (*Cherax quadricarinatus*) // *Agrokompleks*. 2021. Vol. 21(2). P. 48—52. <https://doi.org/10.51978/japp.v21i2.369>.
129. Tampubolon I., Maitindom F. A. Length weight relationship of fresh water lobster (*Cherax quadricarinatus*) in Lake Paniai, Paniai district // *Jurnal Cakrawala Ilmiah*. 2023. Vol. 2(8). P. 3251—3260. <https://doi.org/10.53625/jcijurnalcakrawalailmiah.v2i8.5518>.
130. Taryono Mashar A., Aryasa S. Management policy of invasive species red claw crayfish (*Cherax quadricarinatus*) at Lido Lake, Bogor Regency // In *IOP Conference Series: Earth and Environmental Science*. 2021. Vol. 744. 012090. <https://doi.org/10.1088/1755-1315/744/1/012090>.
131. Tee Z-B., Ibrahim S., Teoh C-Y. Comparative study on the nutritional content and physical attributes of giant freshwater prawn (*Macrobrachium rosenbergii*) and redclaw crayfish (*Cherax quadricarinatus*) meats : preprint (Version 1) available at *Research Square*. 2022. 07 June. <https://doi.org/10.21203/rs.3.rs-1695209/v1>.
132. The chromosome-level genome of



- <https://doi.org/10.18475/cjos.v53i2.a21>.
131. Vecchioni, L., Marrone, F., Chirco, P., Arizza, V., Tricarico, E., & Arculeo M. (2022). An update of the known distribution and status of *Cherax* spp. in Italy (Crustacea, Parastacidae). *BioInvasions Records*, 11(4), 1045–1055. <https://doi.org/10.3391/bir.2022.11.4.22>.
132. Veenstra, J. A. (2020). Gonadulins, the fourth type of insulin-related peptides in decapods. *General and Comparative Endocrinology*, 296, 113528. <https://doi.org/10.1016/j.ygcen.2020.113528>.
133. Wang, Y., Wang, B., Shao, X., Liu, M., Jiang, K., Wang, M., & Wang, L. (2020). A comparative transcriptomic analysis in late embryogenesis of the red claw crayfish *Cherax quadricarinatus*. *Molecular genetics and genomics*, 295(2), 299–311. <https://doi.org/10.1007/s00438-019-01621-4>.
134. Wang, Y., Wang, B., Shao, X., Liu, M., Jiang, K., Wang, M., & Wang, L. (2020). Identification and profiling of MicroRNAs during embryogenesis in the red claw crayfish *Cherax quadricarinatus*. *Frontiers in physiology*, 11, 878. <https://doi.org/10.3389/fphys.2020.00878>.
135. Wang, Y., Wang, B.J., Liu, M., Jiang, K.Y., Wang, M.Q., & Wang, L. (2020). The first identification of a C-type lectin gene (CqCTL) in *Cherax quadricarinatus*: sequence features and expression profiles. *Invertebrate Survival Journal*, 17(1), 108–116. <https://doi.org/10.25431/1824-307X/isj.v0i0.108-116>.
136. Wei, M., Gu, Z.F., Pan, Z., Shi, Y. H., Huang, Z. W., Zheng, X., Liao, X. R., Li, J. N., Liu, C. S., & Wang, A. M. (2020). Effects of background color on growth, survival, body color, and inhabiting behavior distribution of *Cherax quadricarinatus*. *Journal of Marine Sciences*, 44, 60–65.
137. Wei, M., Wang, A., Gu, Z., Liu, C., Li, J., Liao, X., & Pan, Z. (2021). Comparison of growth performance, carotenoid *Cherax quadricarinatus* / Chen H. et al. // Scientific data. 2023. Vol. 10(1). P. 215. <https://doi.org/10.1038/s41597-023-02124-z>.
133. The combined effect of vitamin E, arachidonic acid, *Haemtococcus pluvialis*, nucleotides and yeast extract on growth and ovarian development of crayfish (*Cherax quadricarinatus*) by the orthogonal array design / Shehata A. I. et al. // Aquaculture Nutrition. 2020. Vol. 26(6). P. 2007—2022. <https://doi.org/10.1111/anu.13142>.
134. The diet of an invasive crayfish, *Cherax quadricarinatus* (Von Martens, 1868), in Lake Kariba, inferred using stomach content and stable isotope analyses / Marufu, L. T. et al. // BioInvasions Records. 2018. Vol. 7(2). P. 121—132. <https://doi.org/10.3391/bir.2018.7.2.03>
135. The effect of the lectin from *Cherax quadricarinatus* on its granular hemocytes / Sánchez-Salgado J. L. et al. // Fish & Shellfish Immunology. 2018. Vol. 77. P. 131—138. <https://doi.org/10.1016/j.fsi.2018.03.050>.
136. The first identification of a C-type lectin gene (CqCTL) in *Cherax quadricarinatus*: sequence features and expression profiles / Wang Y. et al. // Invertebrate Survival Journal. 2020. Vol. 17(1). P. 108—116. <https://doi.org/10.25431/1824-307X/isj.v0i0.108-116>.
137. The first occurrence of the Australian redclaw crayfish *Cherax quadricarinatus* (von Martens, 1868) in the contiguous United States / Morningstar C. R. et al. // BioInvasions Records. 2020. Vol. 9(1). P. 120—126. <https://doi.org/10.3391/bir.2020.9.1.16>.
138. The introduction, spread and ecology of redclaw crayfish *Cherax quadri-*



- content, and temperature tolerance of two-colored strains of the red claw crayfish *Cherax quadricarinatus*. *Journal of Shellfish Research*, 40(2), 421–427. <https://doi.org/10.2983/035.040.0214>.
138. Wei, Y., Cheng, S., Jia, Y., Chi, M., Liu, S., Zheng, J., & Gu, Z. (2022). Effects of nitrite on physiological indices and intestinal flora of parent *Cherax quadricarinatus* during overwintering. *Journal of Shanghai Ocean University*, 31(1), 278–287. <https://doi.org/10.12024/jso.20201203256>.
139. Weiperth A., Gál B., Kuříková P., Langrová I., Kouba A., & Patoka J. (2019). Risk assessment of pet-traded decapod crustaceans in Hungary with evidence of *Cherax quadricarinatus* (von Martens, 1868) in the wild. *North-Western Journal of Zoology*, 15(1), 42–47.
140. Wen-ping, J., Shun, C., Yong-yi, J., Mei-li, C., Shi-li, L., Jian-bo, Z., Fei, Li., Yi-nuo, Li., Zhi-min, G., & Dan-li, W. (2022). A study on the effects of light intensity on juveniles of the red claw crayfish *Cherax quadricarinatus*. *Aquaculture Research*, 53(18), 6454–6462. <https://doi.org/10.1111/are.16115>.
141. Weyl, O. L. F., Nunes, A. L., Ellender, B. R., Weyl, P. S. R., Chilala, A. C., Jacobs, F. J., Murray-Hudson, M., & Douthwaite, R. J. (2017). Why suggesting Australian redclaw crayfish *Cherax quadricarinatus* as biological control agents for snails is a bad idea. *African Journal of Aquatic Science*, 42(4), 325–327. <https://doi.org/10.2989/16085914.2017.1414685>.
142. Wu, D. L., Rao, Q. X., Cheng, L., Lv, W. W., Zhao, Y. L., & Song, W. G. (2021). Cloning and characterisation of a $\Delta 9$ fatty acyl desaturase-like gene from the red claw crayfish (*Cherax quadricarinatus*) and its expression analysis under cold stress. *Journal of Thermal Biology*, 102, 103122. <https://doi.org/10.1016/j.jtherbio.2021.103122>.
- carinatus* in the Zambezi catchment / Douthwaite R. J. et al. // *African Journal of Aquatic Science*. 2018. Vol. 43. P. 353–366. <https://doi.org/10.2989/16085914.2018.1517080>.
139. The invasive crayfish *Cherax quadricarinatus* facing chlordecone in Martinique: bioaccumulation and depuration study / Baudry T. et al. // *Chemosphere*. 2022. Vol. 286. 131926. <https://doi.org/10.1016/j.chemosphere.2021.131926>.
140. The survival rate and growth of juvenile crayfish (*Cherax quadricarinatus*) with different types of shelter / Mamuya J. et al. // *Jurnal Ilmiah Platax*. 2019. Vol. 7(2). P. 427–431. <http://dx.doi.org/10.35800/jip.7.2.2019.24510>.
141. The use of nutmeg oil (*Myristica fragrans*) Houttuyn 1774 as anesthetic component in different concentrations for the young freshwater lobster (*Cherax quadricarinatus*) Von Martens 1868 / Mamuko N. et al. // *E-Journal Budidaya Perairan*. 2021. Vol. 10(1). P. 73–80. <https://doi.org/10.35800/bdp.10.1.2022.35535>.
142. Tierney L. J., Wild C. H., Furse J. M. Total incombustible (mineral) content of *Cherax quadricarinatus* differs between feral populations in Central-Eastern Australia // *PeerJ*. 2019. Vol. 17. <http://dx.doi.org/10.7717/peerj.6351>.
143. Tiftazani M. H., Suparmi Sumarto. Storage time of freshwater lobster (*Cherax quadricarinatus*) with green betel leaf anesthesia (*Piper betle* L.) // *Terubuk*. 2022. Vol. 50(3). P. 1619–1623.
144. Todd S.-R. L., Hyslop E. J. Morphometrics, diet, reproductive biology, and ecological interactions of the introduced redclaw crayfish, *Cherax quadricarinatus* (Decapoda: Parastacidae), in Jamaic



143. Wu, D., Liu, Z., Yu, P., Huang, Y., Cai, M., Zhang, M., & Zhao, Y. (2020). Cold stress regulates lipid metabolism via AMPK signalling in *Cherax quadricarinatus*. *Journal of thermal biology*, 92, 102693. <https://doi.org/10.1016/j.jtherbio.2020.102693>.
144. Xu, X., Duan, H., Shi, Y., Xie, S., Song, Z., Jin, S., Li F., & Xiang, J. (2018). Development of a primary culture system for haematopoietic tissue cells from *Cherax quadricarinatus* and an exploration of transfection methods. *Developmental and Comparative Immunology*, 88, 45–54. <https://doi.org/10.1016/j.dci.2018.07.006>.
145. Xu, Y., Pan, H., Lu, M., Liu, Q., Shafique, L., Peng, J., Ahmed T., Wang R., Zhang H., Wang Q., & Zhu, P. (2021). Analysis of transcripts and splice isoforms in red claw crayfish (*Cherax quadricarinatus*) using single-molecule long-read sequencing. *Aquaculture*, 541, 736828. <https://doi.org/10.1016/j.aquaculture.2021.736828>.
146. Yau, S. M., & Lau, A. (2021). First record of the australian redclaw crayfish *Cherax quadricarinatus* (Von Martens, 1868) in Hong Kong, China. *BioInvasions Records*, 10(2), 369–377. <https://doi.org/10.3391/bir.2021.10.2.15>
147. Yuan, J., Guo, J., Wang, H., Guo, A., Lian, Q., & Gu, Z. (2019). Acute toxicity of cypermethrin on the juvenile of red claw crayfish *Cherax quadricarinatus*. *Chemosphere*, 237, 124468. <https://doi.org/10.1016/j.chemosphere.2019.124468>.
148. Yuan, J., Zheng, Y., & Gu, Z. (2021). Effects of cypermethrin on the hepatic transcriptome and proteome of the red claw crayfish *Cherax quadricarinatus*. *Chemosphere*, 263, 128060. <https://doi.org/10.1016/j.chemosphere.2020.128060>.
149. Zheng, J., Cai, L., Jia, Y., Chi, M., Cheng, S., Liu, S., Li, F., & Gu, Z. ca, West Indies // *Caribbean Journal of Science*. 2023. Vol. 53(2). P. 397—410. <https://doi.org/10.18475/cjos.v53i2.a21>.
145. Transcriptomic analysis of crustacean molting gland (Y-organ) regulation via the mTOR signaling pathway / Shyamal S. et al. // *Scientific Reports*. 2018. Vol. 8. 7307. <https://doi.org/10.1038/s41598-018-25368-x>.
146. Two homogametic genotypes — one crayfish: on the consequences of intersexuality / Levy T. et al. // *iScience*. 2020. Vol. 23. 101652. <https://doi.org/10.1016/j.isci.2020.101652>.
147. Veenstra J. A. Gonadulins, the fourth type of insulin-related peptides in decapods // *General and Comparative Endocrinology*. 2020. Vol. 296. 113528. <https://doi.org/10.1016/j.ygcen.2020.113528>.
148. Why suggesting australian redclaw crayfish *Cherax quadricarinatus* as biological control agents for snails is a bad idea / Weyl O. L. F. et al. // *African Journal of Aquatic Science*. 2017. Vol. 42(4). P. 325—327. <https://doi.org/10.2989/16085914.2017.1414685>.
149. Yau S. M., Lau A. First record of the australian redclaw crayfish *Cherax quadricarinatus* (Von Martens, 1868) in Hong Kong, China // *BioInvasions Records*. 2021. Vol. 10(2). P. 369—377. <https://doi.org/10.3391/bir.2021.10.2.15>.
150. Yuan J., Zheng Y., Gu Z. Effects of cypermethrin on the hepatic transcriptome and proteome of the red claw crayfish *Cherax quadricarinatus* // *Chemosphere*. 2021. Vol. 263. 128060. <https://doi.org/10.1016/j.chemosphere.2020.128060>.
151. Zhu K., Yang F., Li F. Molecular markers for hemocyte subpopulations



- (2020). Identification and functional analysis of the doublesex gene in the redclaw crayfish, *Cherax quadricarinatus*. *Gene expression patterns*, 37, 119129. <https://doi.org/10.1016/j.gep.2020.119129>.
150. Zheng, J., Chen, L., Jia, Y., Chi, M., Li, F., Cheng, S., Liu, S., Liu, Y., & Gu Z. (2022). Genomic structure, expression, and functional characterization of the Fem-1 gene family in the redclaw crayfish, *Cherax quadricarinatus*. *General and comparative endocrinology*, 316, 113961. <https://doi.org/10.1016/j.ygcen.2021.113961>.
151. Zheng, J., Cheng, S., Jia, Y., Gu, Z., Li, F., Chi, M., Liu, S., & Jiang, W. (2019). Molecular identification and expression profiles of four splice variants of sex-lethal gene in *Cherax quadricarinatus*. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 234, 26–33. <https://doi.org/10.1016/j.cbpb.2019.05.002>.
152. Zheng-Bin, T., Saadiah, I., & Chaiw-Yee, T. (2022). Comparative study on the nutritional content and physical attributes of giant freshwater prawn (*Macrobrachium rosenbergii*) and redclaw crayfish (*Cherax quadricarinatus*) meats: preprint (Version 1) 07 June. *Research Square*. <https://doi.org/10.21203/rs.3.rs-1695209/v1>.
153. Zhou, S., Jiang, K., Liu, M., Wang, B., & Wang, L. (2022). Exploration of the functional properties of red cuticle patch of redclaw crayfish (*Cherax quadricarinatus*) by histological and transcriptomic analysis. *Aquaculture*, 561(15), 738624. <https://doi.org/10.1016/j.aquaculture.2022.738624>.
154. Zhu, K., Yang, F., & Li, F. (2022). Molecular markers for hemocyte subpopulations in crayfish *Cherax quadricarinatus*. *Developmental and comparative immunology*, 132, 104407. <https://doi.org/10.1016/j.dci.2022.104407>.
- in crayfish *Cherax quadricarinatus* // Developmental and comparative immunology. 2022. Vol. 132. 104407. <https://doi.org/10.1016/j.dci.2022.104407>.
152. Зв'язок хімічних та фізичних показників води з морфологічними ознаками раків різних видів / Федорович Є. І. та ін. // Вісник Сумського національного аграрного університету. 2022. № 4(47). С. 165—170. <https://doi.org/10.32845/bsnau.lvst.2021.4.28>.
153. Особливості зовнішньої будови *Cherax quadricarinatus* (Von Martens, 1868) / Гриневич Н. Є. та ін. // Problems of science and practice, tasks and ways to solve them : XX International scientific and practical conference, Warsaw, Poland, May 24-27 2022 : proceed. Warsaw, 2022. P. 44—46.
154. Особливості процесу линьки раків різних видів / Федорович Є. І. та ін. // Таврійський науковий вісник. 2022. Вип. 126. С. 230—237. <https://doi.org/10.32851/2226-0099.2022.126.32>.

